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Challenges in AMR *K. pneumoniae* infections and treatment,

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Utstein Kloster, 3 May 2018

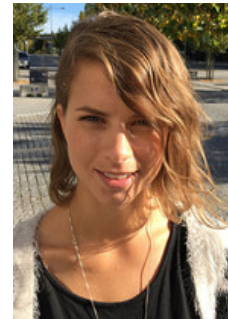
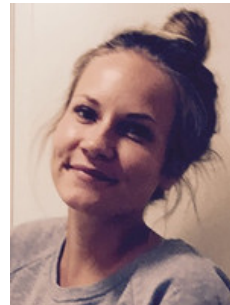


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Research group



Department of Laboratory Medicine



Karolinska University Hospital

<http://ki.se/en/labmed/christian-giske-group>

Funding



Vetenskapsrådet



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JOINT PROGRAMMING
INITIATIVE ON ANTIMICROBIAL
RESISTANCE



Svenska
Läkaresällskapet

When does antimicrobial resistance really matter?

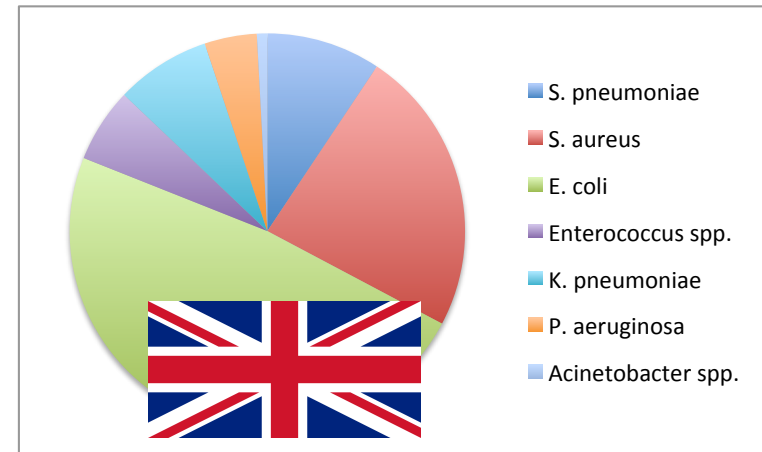
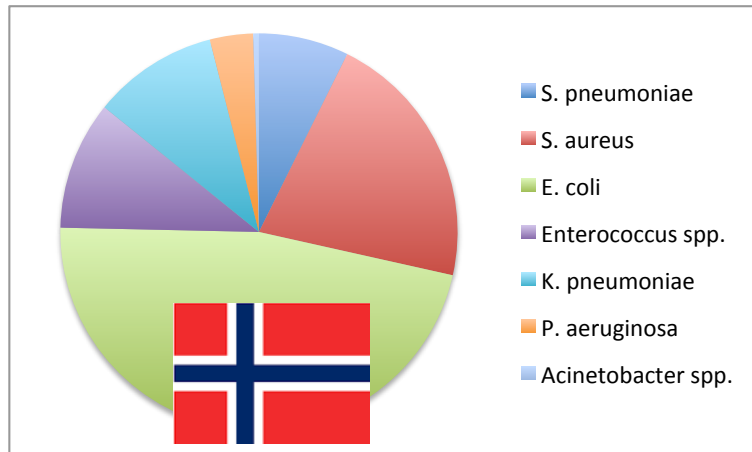
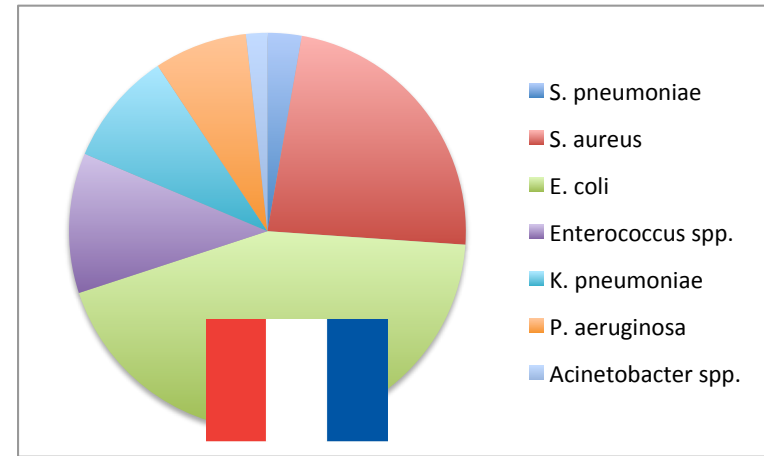
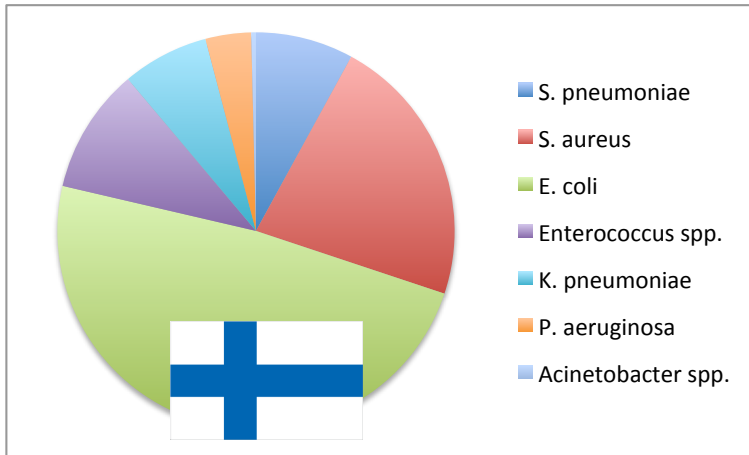
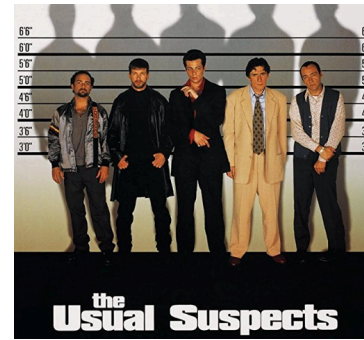
- In severe disease: as a cause of mortality
 - Frequently difficult to ascertain contribution to death
 - Patient comorbidities can be a significant bias
 - Some types of resistance come with greatly reduced fitness
 - The role of strain factors usually insufficiently studied
 - In other diseases: only when using “soft parameters”
 - Length of stay/increased costs
 - Impact on strain transmission
 - Long-term consequences
 - Bloodstream infections: undisputable relevance, frequently access to strains, easier to compare between countries
-

In vitro resistance vs clinical response

- Rex and Pfaller 2002 (antifungal AST)
 - Susceptible=responds in 90% of the cases
 - Resistant=still responds in around 60% of the cases
 - Pertains to immunocompetent individuals with monomicrobial infections with predictable penetration of the drug to the infection site
 - Polymicrobial infections with unpredictable penetration: even lower
- Strain variations
 - Are usually major in preclinical PK-PD data
 - Strain virulence is not taken into account in AST
 - We also do not take into account individual variation in the immune system

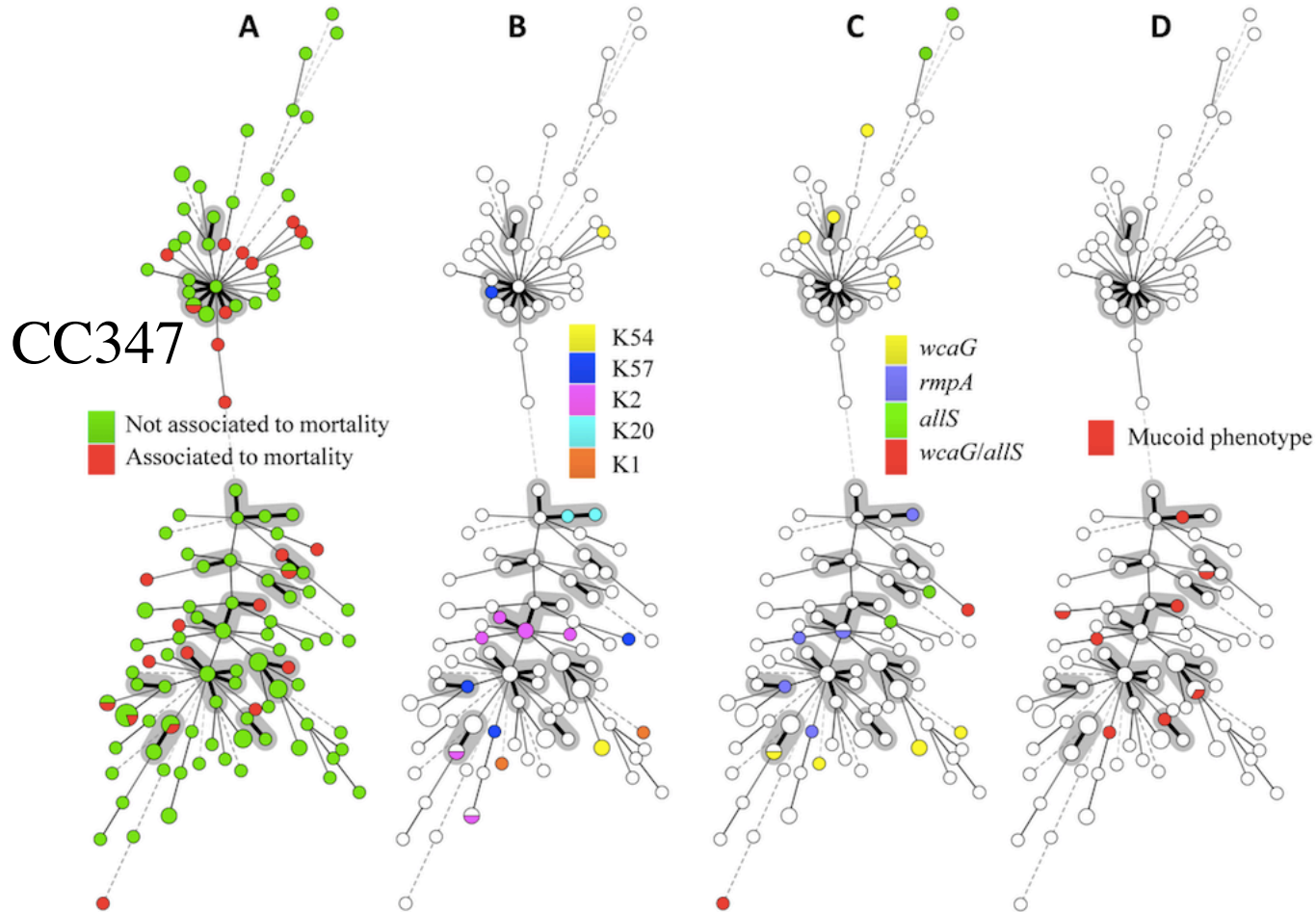
Doern GV and Brecher SM. JCM 2012

Bloodstream infections:



Klebsiella pneumoniae – invasive infections

K. pneumoniae (sensu lato), invasive infections Stockholm



Maatallah M et al. PloS One 2014. ST380, K2, *rmpA* was encountered, but was not associated with a fatal outcome

K. pneumoniae (sensu lato), invasive infections Stockholm

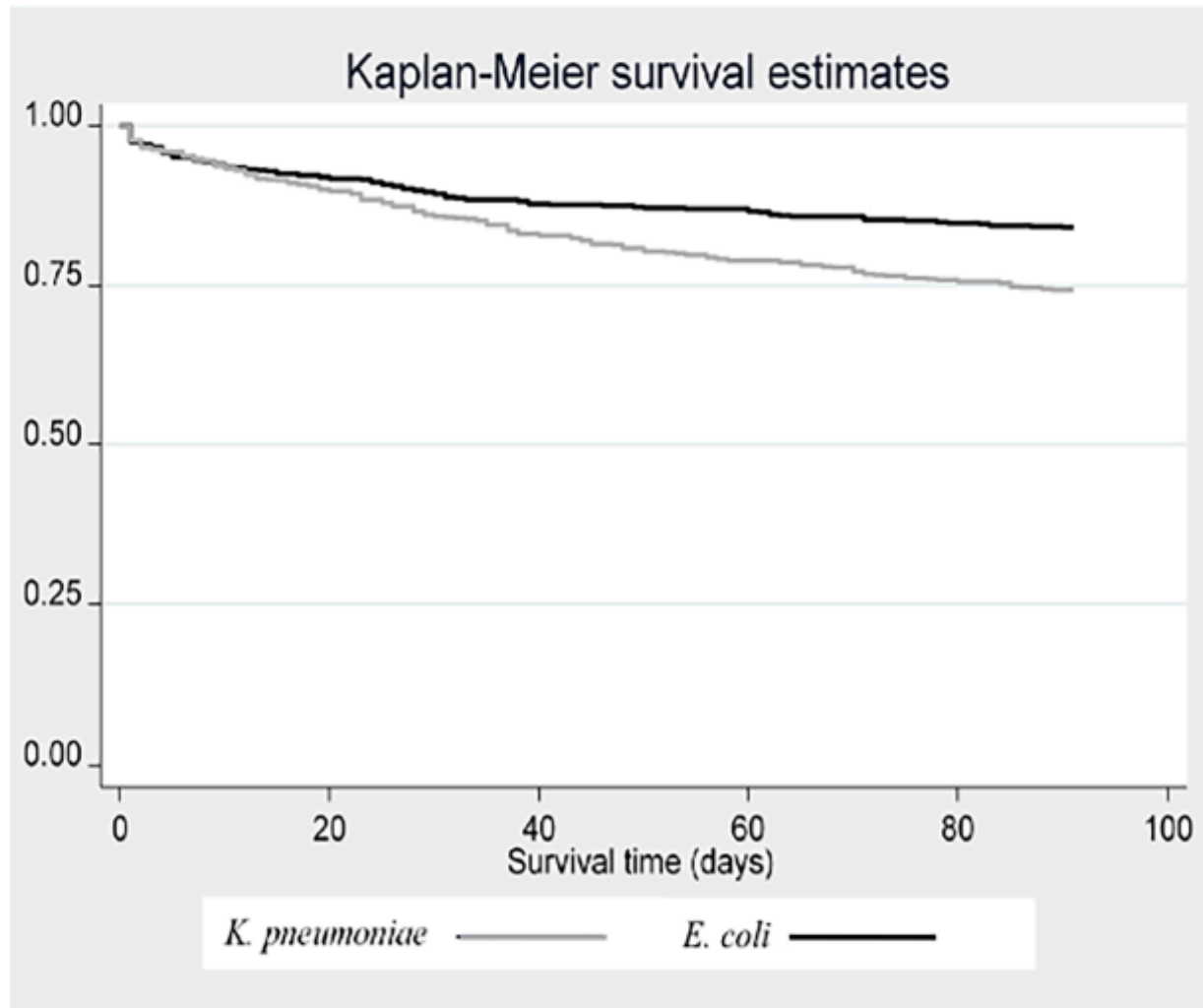
Parameter	Dead within 30 d	Survivors	P-value
Charlson (median)	5	2	0.0003
Metastatic cancer	43.5%	11.2%	0.001
KpIII (<i>variicola</i>)	43.5%	22.4%	

Invasive infection of *K. pneumoniae* vs *E. coli*

Table 1. Clinical characteristics of patients with invasive infection caused by *K. pneumoniae* versus *E. coli*, multivariable analysis.

	<i>K. pneumoniae</i> n = 599	<i>E. coli</i> n = 599	Adjusted odds ratio (95% CI)
<i>Patients factors</i>	No (%)	No (%)	<i>K. pneumoniae</i> vs <i>E. coli</i>
Peripheral vascular disease	30 (5)	14 (2)	3.74 (1.65–8.48)
COPD	58 (10)	37 (6)	1.96 (1.14–3.36)
Kidney disease	105 (18)	69 (12)	1.90 (1.28–2.82)
Bile disease	36 (6)	15 (3)	3.10 (1.44–6.66)
Hematological malignancy	112 (19)	76 (13)	1.70 (1.07–2.70)
Bile/liver/pancreas malignancy	51 (9)	22 (4)	3.45 (1.77–6.75)
Colorectal malignancy	42 (7)	24 (4)	2.56 (1.34–4.89)
Urinary catheter	191 (32)	111 (19)	2.36 (1.64–3.40)
Central catheter	190 (32)	96 (16)	2.32 (1.53–3.54)
Hospital-acquired ^{a)}	178 (30)	197 (33)	0.53 (0.37–0.77)
Healthcare- associated community-onset ^{a)}	163 (27)	55 (9)	3.06 (2.03–4.62)

Higher 30-d mortality in *K. pneumoniae*



P<0.001

Factors associated with mortality

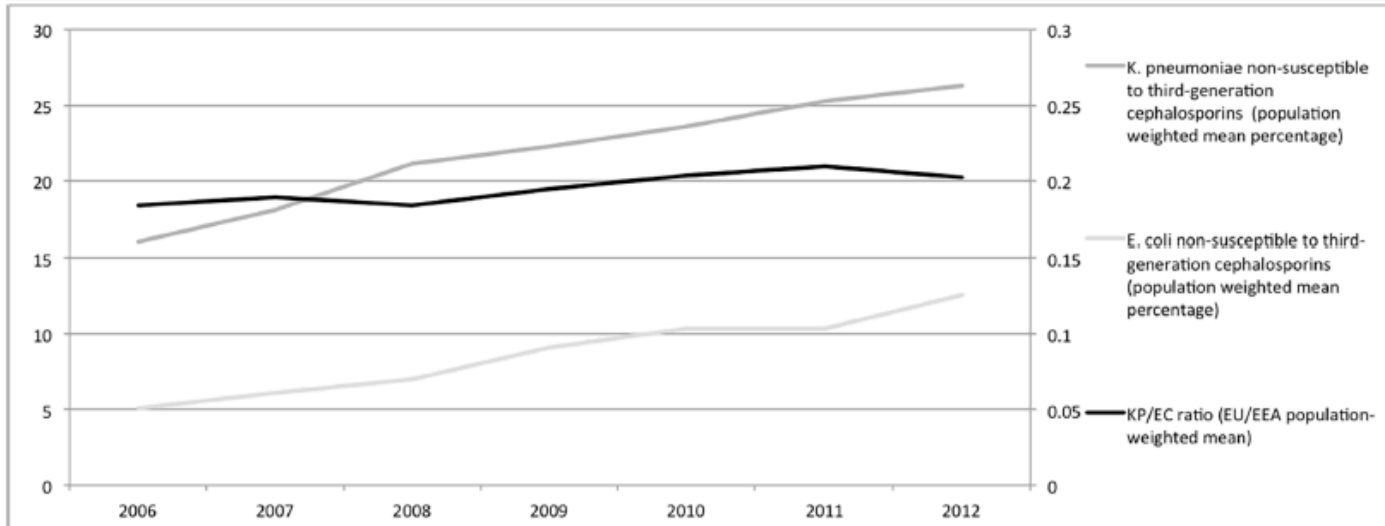
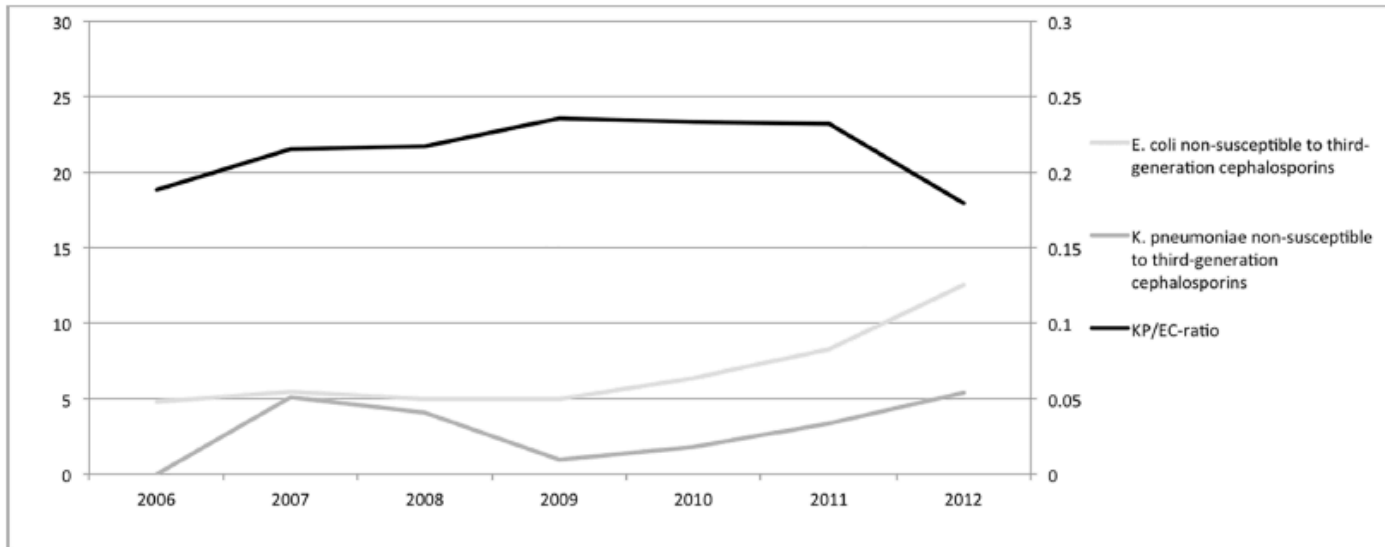
Table 2. Associated factors for mortality in the extended *K. pneumoniae* cohort, factors significant in multivariable analysis.

	Mortality within 7d (n = 43)	Mortality within 30 d (n = 101)	Mortality within 90 d (n = 176)
<i>Associated factors*</i>	Adjusted OR	Adjusted OR	Adjusted OR
Age	1.03 (1.00–1.05)	1.02 (1.00–1.04)	1.03 (1.01–1.04)
Polymicrobial infection	3.07 (1.51–6.27)	2.20 (1.32–3.68)	
Kidney disease			2.33 (1.41–3.84)
CNS disease		3.12 (1.73–5.62)	2.09 (1.26–3.45)
Lung malignancy	13.45 (3.94–45.90)	13.20 (4.11–42.38)	20.77 (6.01–71.73)
Urogenital, GI, bile/liver/pancreas malignancy		2.07 (1.10–3.91)	3.07 (1.87–5.05)
Hematological malignancy		3.13 (1.47–6.63)	2.50 (1.33–4.72)
Other malignancy**	6.18 (1.87–20.41)	6.34 (2.54–15.85)	3.77 (1.63–8.74)
Hospital-acquired			1.99 (1.21–3.28)
Healthcare-associated community-onset			1.69 (1.02–2.79)
<i>Source of infection</i>			
Respiratory tract	3.62 (1.01–13.04)	3.79 (1.32–10.87)	3.74 (1.44–9.68)
Bile/liver, GI		1.92 (1.00–4.16)	1.91 (1.15–3.15)
Unknown		2.09 (1.05–4.16)	

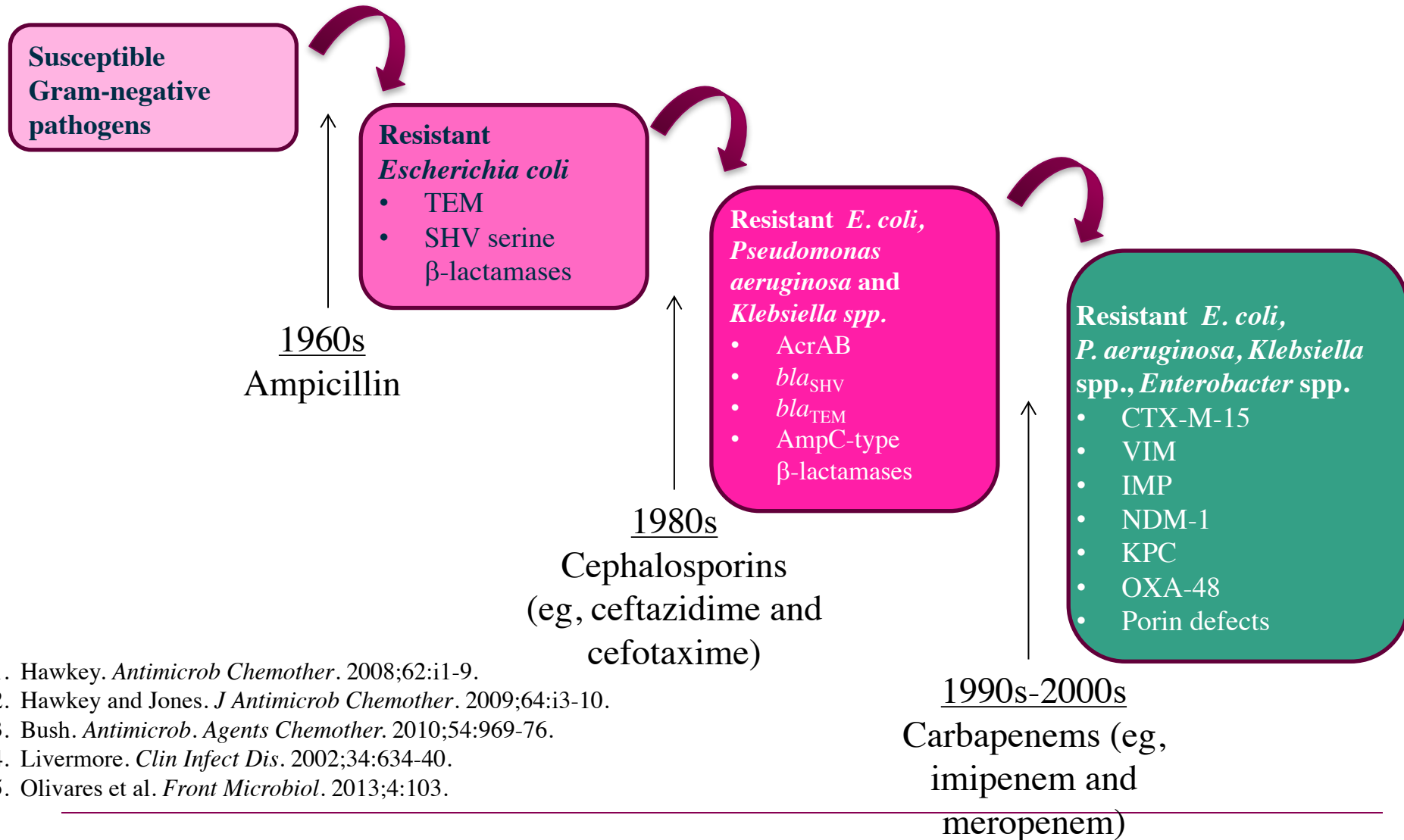
K. pneumoniae: expanding pathogen in Europe, not in Sweden



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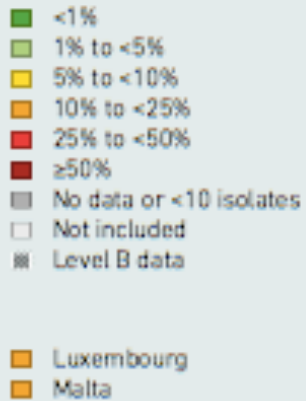


Evolution of Gram-negative Resistance

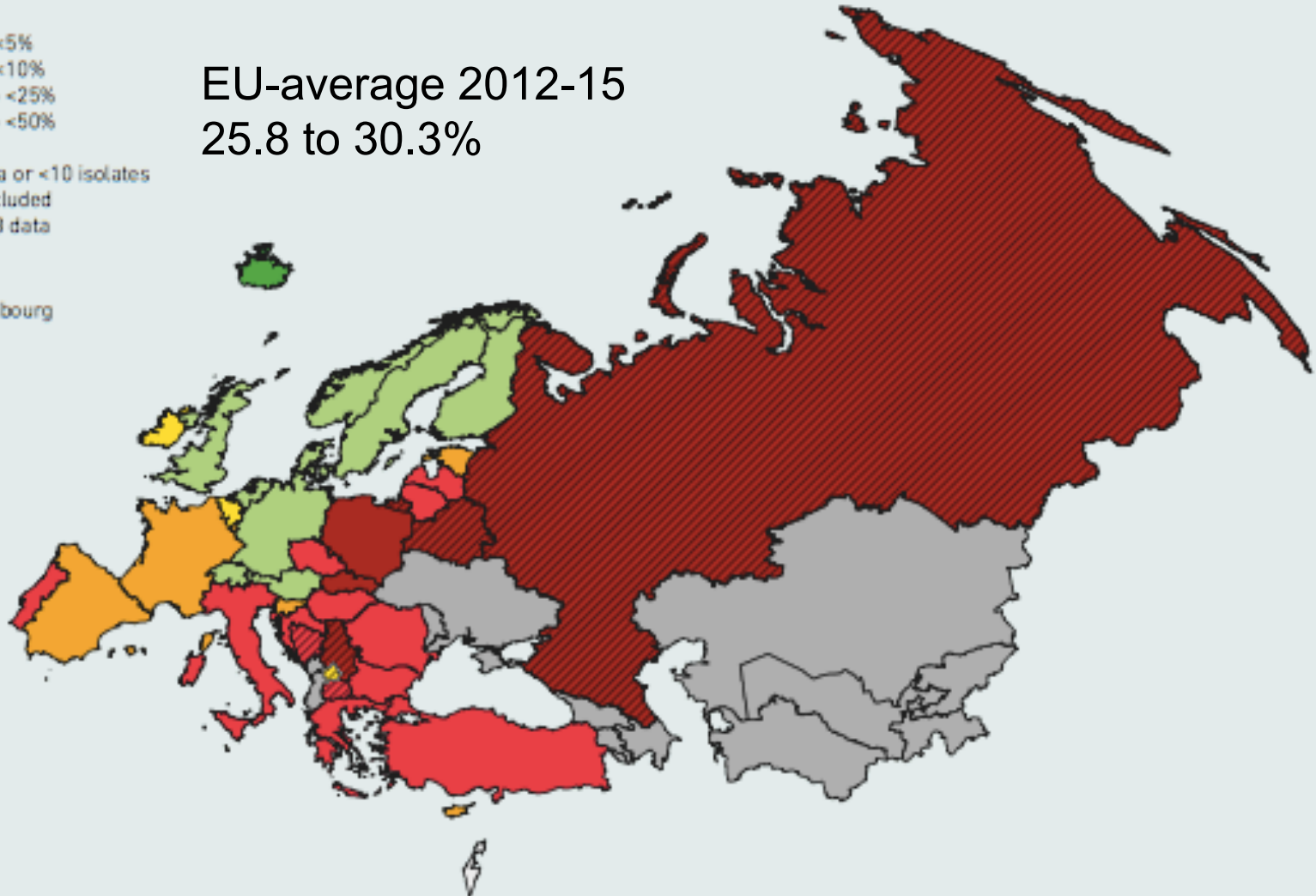


1. Hawkey. *Antimicrob Chemother.* 2008;62:i1-9.
2. Hawkey and Jones. *J Antimicrob Chemother.* 2009;64:i3-10.
3. Bush. *Antimicrob. Agents Chemother.* 2010;54:969-76.
4. Livermore. *Clin Infect Dis.* 2002;34:634-40.
5. Olivares et al. *Front Microbiol.* 2013;4:103.

ESBL-producing *K. pneumoniae*



EU-average 2012-15
25.8 to 30.3%

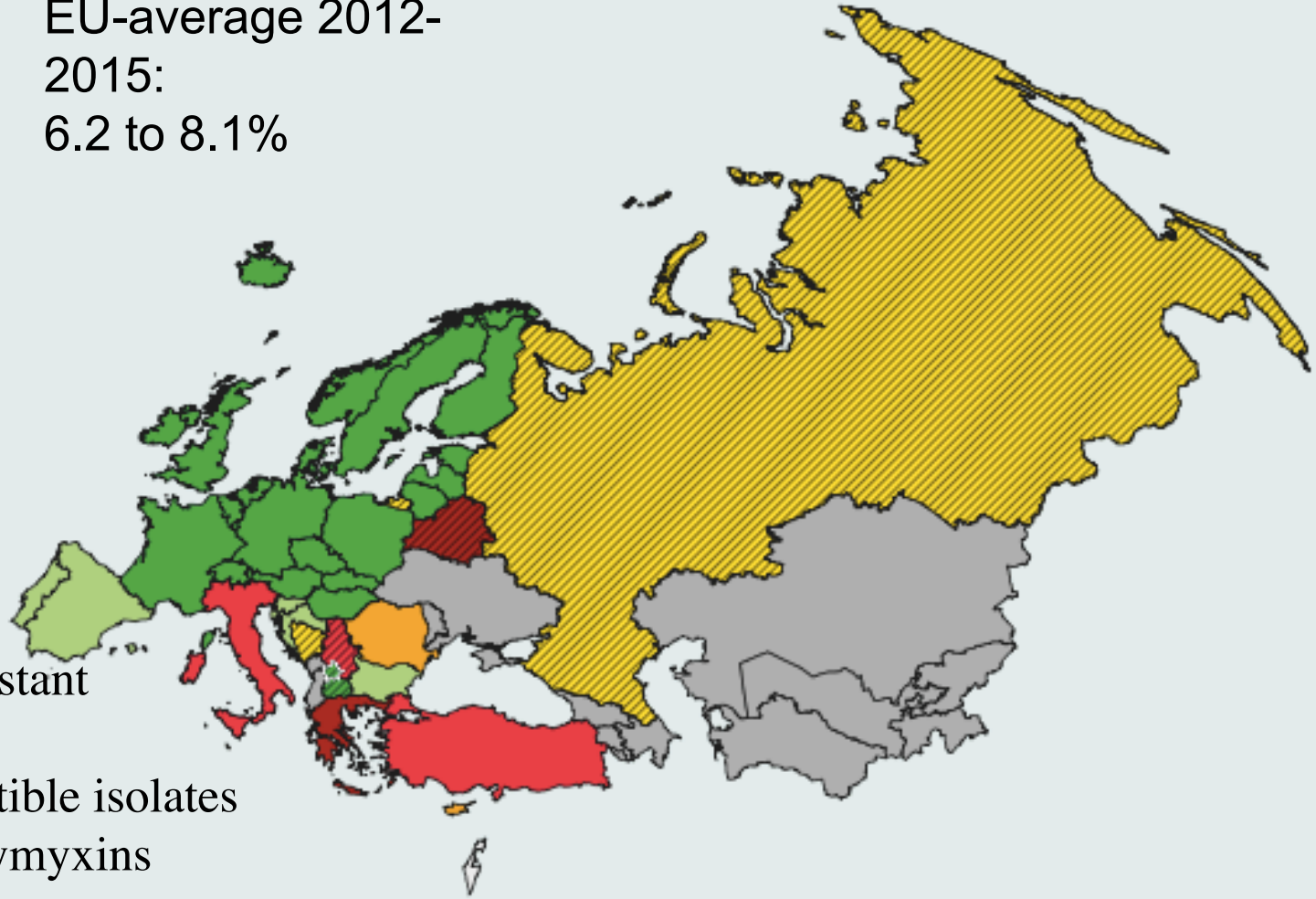


Carbapenem resistant *K. pneumoniae*

- <1%
- 1% to <5%
- 5% to <10%
- 10% to <25%
- 25% to <50%
- ≥50%
- No data or <10 isolates
- Not included
- Level B data

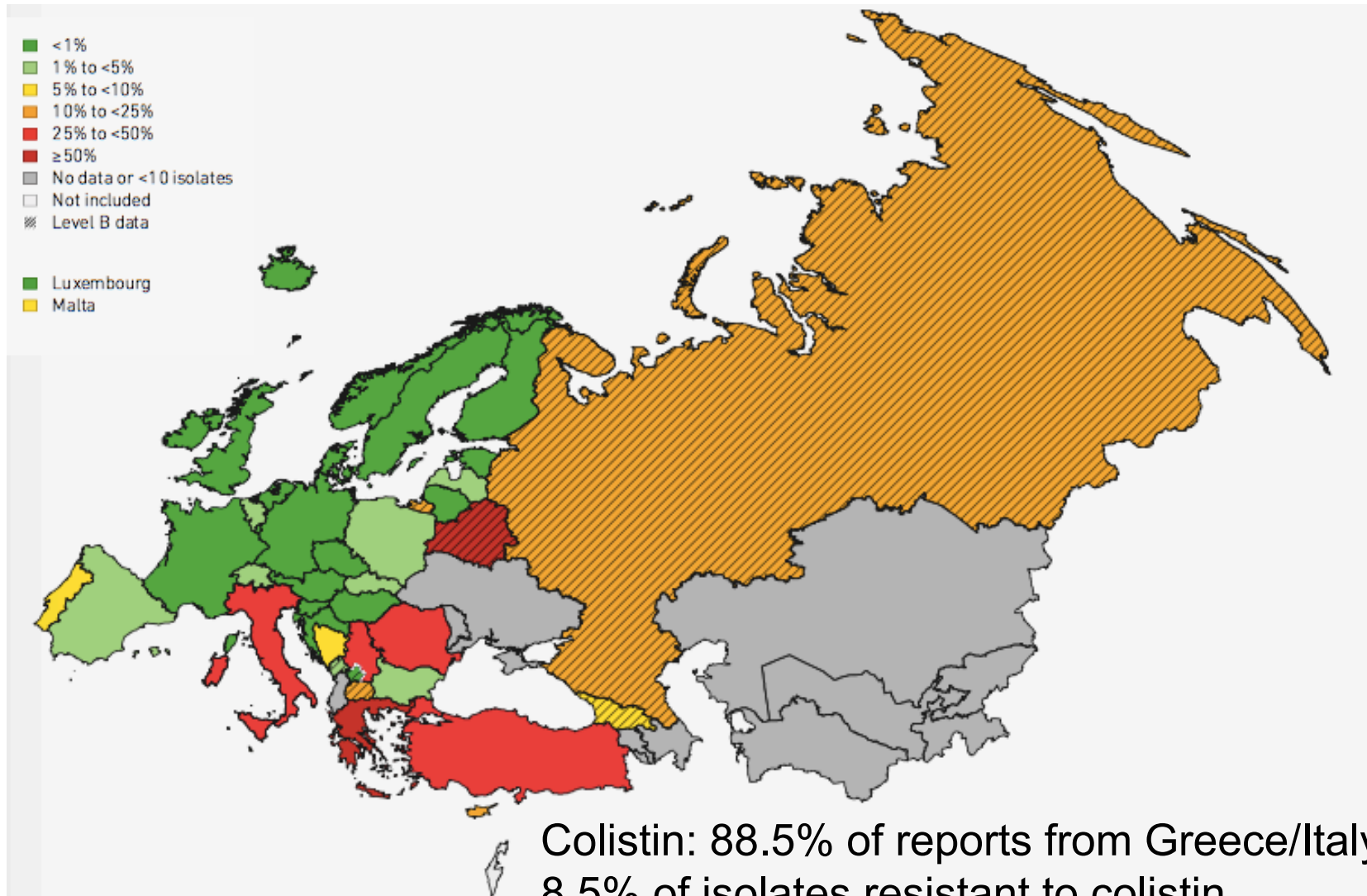
EU-average 2012-
2015:
6.2 to 8.1%

- Luxembourg
- Malta

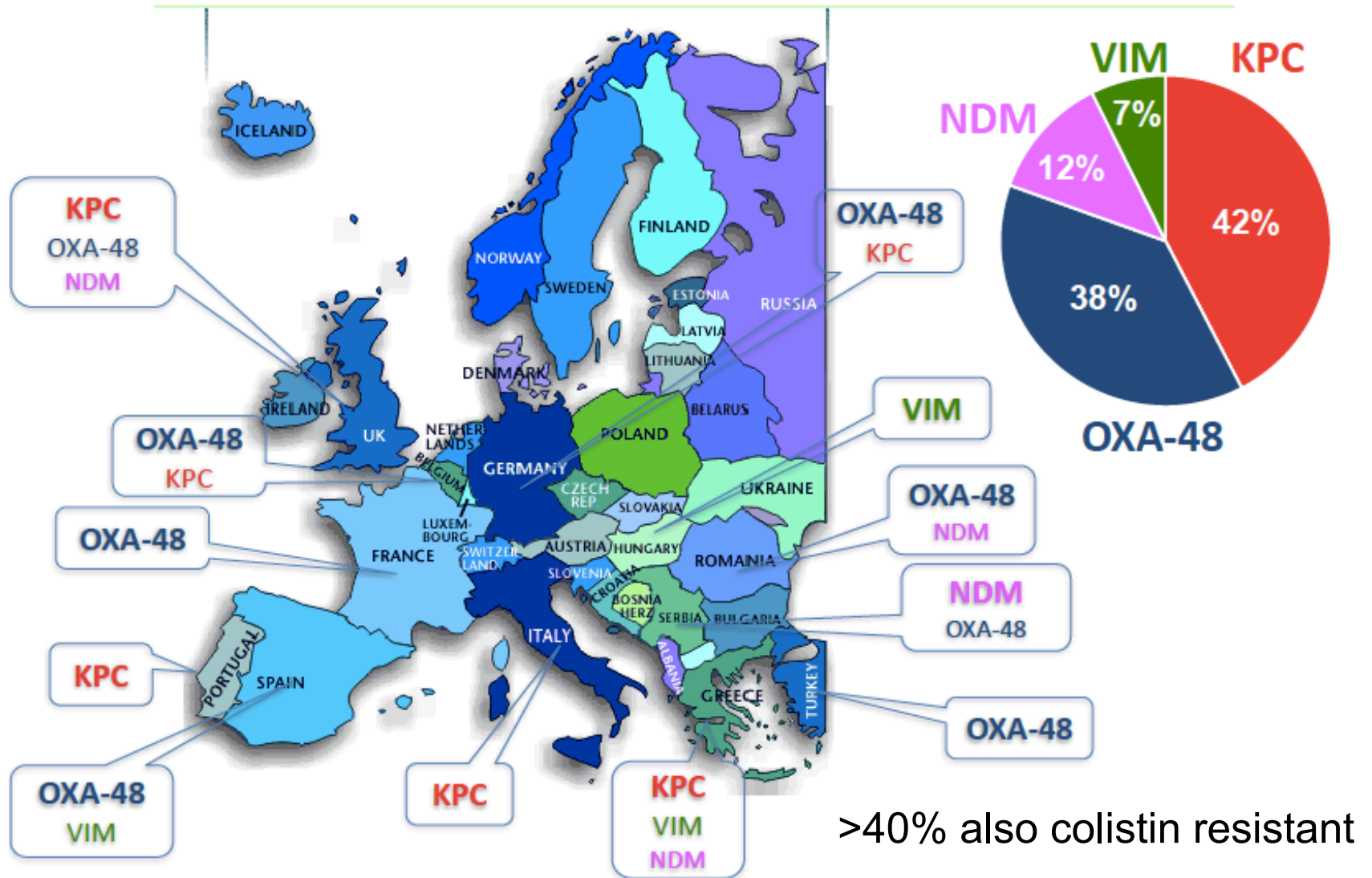


31.9% also resistant
to polymyxins
2.6% of susceptible isolates
resistant to polymyxins

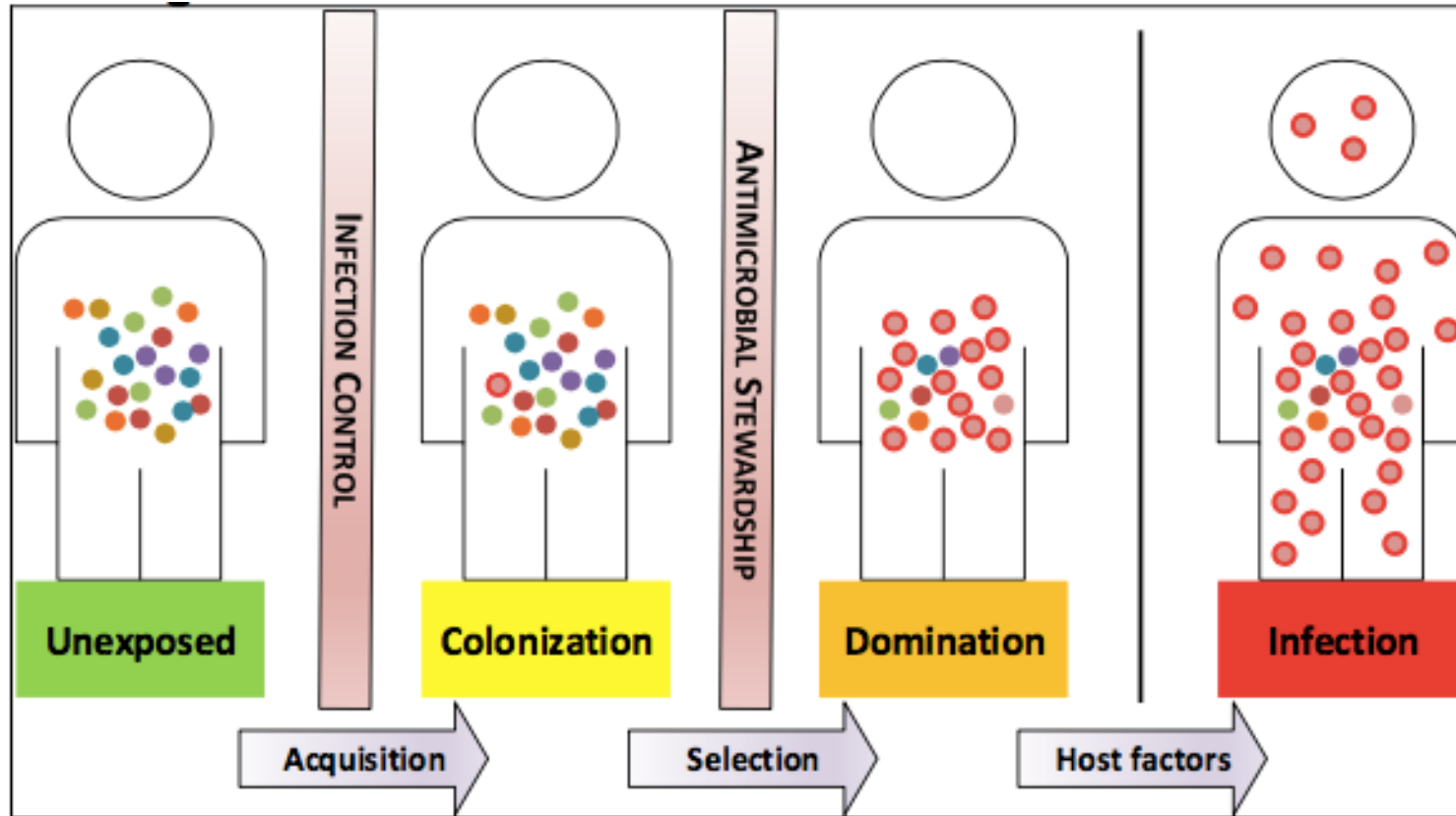
Carbapenem-R *K. pneumoniae*, 2016



CPE genotypes – situation in Europe

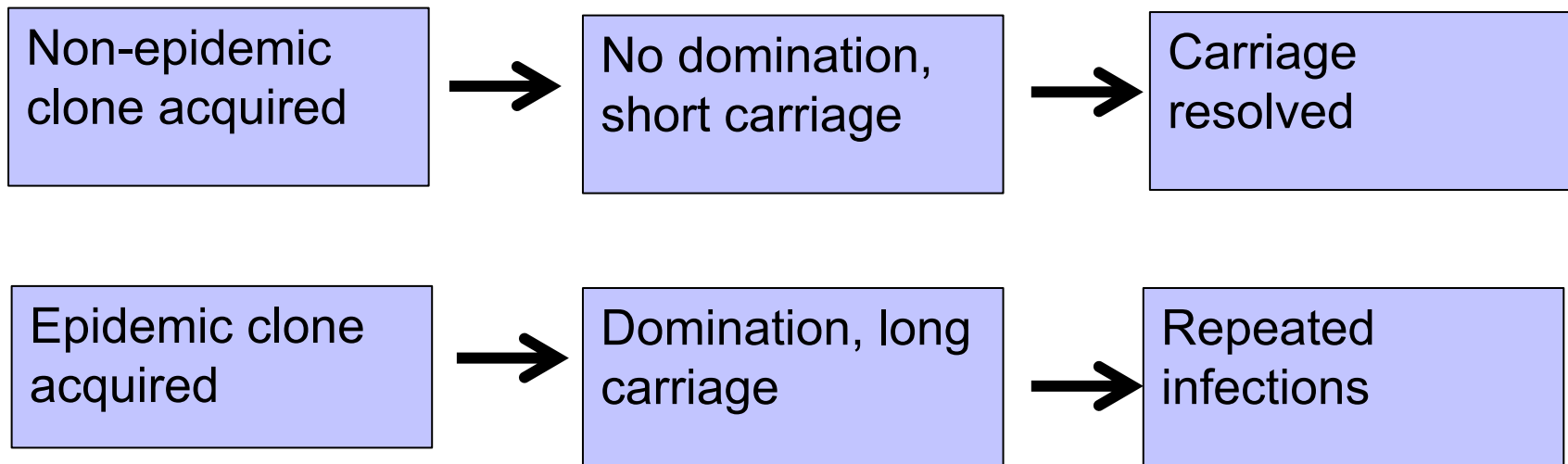


The process from colonization to infection



Hypothesis under investigation in recently funded multicentre study (Joint Programming Initiative AMR, JPIAMR)

Colonization to infection – a hypothesis



- Process likely influenced by antimicrobial consumption
 - We have the tools to stratify in high-risk and low-risk carriage
 - Therapeutic interventions: directed towards high-risk carriage
 - Effects could be monitored by continuous surveillance of clones occurring in bloodstream infection
-

Klebsiella pneumoniae – novel treatment

Activity of new drugs

Antimicrobial	EBA ESBL	EBA AmpC	KPC KP	EBA MBL	PA MDR	AB MDR	SM
Ceftolozane-tazobactam	+	-	-	-	+	-	-
Ceftazidime-avibactam	+	+	+	-	+/-	-	-
Meropenem-vaborbactam	+	+	+	-	+/-	-	-
Imipenem-relebactam	+	+	+	-	+/-	-	-
Cefiderocol	+	+	+	+	+	+	+
Plazomicin	+	+	+	+/-	+	+	-
Eravacycline	+(?)	+(?)	+(?)	+(?)	-	?	?
Colistin	+	+	+	+	+	+	+

Adapted from Falagas ME et al. Expert Review of Anti-infect. Ther. 2016; 8:747-763

Emerging resistance to new agents...

- *K. pneumoniae* ST258, bloodstream infection – isolates on day 1 (MIC 4 mg/L) and 2 (MIC 32 mg/L) (meropenem therapy)
- PacBio sequencing of both strains
- OmpK35 and K36 mutations
- Increase expression of KPC-3 (multiple copies of Tn4401 transposon containing KPC-3)

Resistance to Ceftazidime-Avibactam in *Klebsiella pneumoniae* Due to Porin Mutations and the Increased Expression of KPC-3

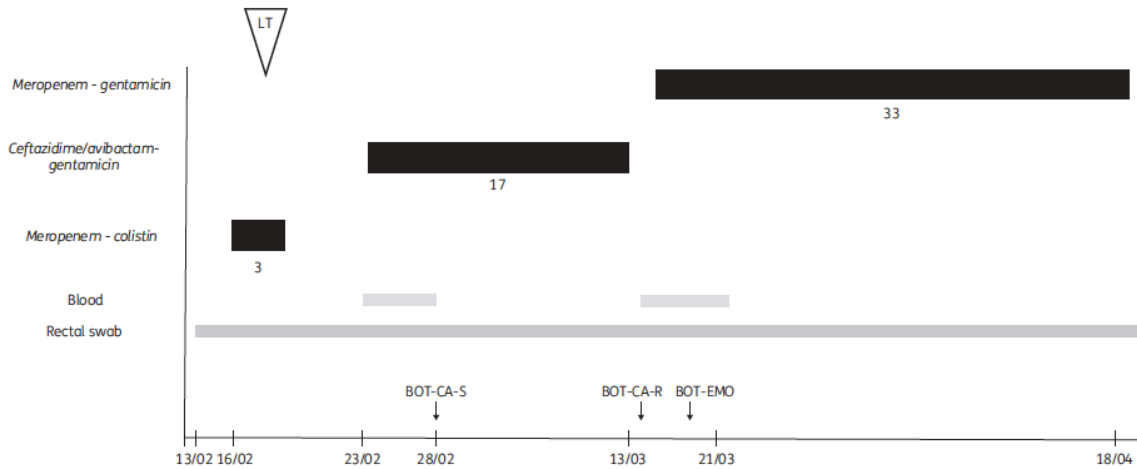
Romney M. Humphries,  Peera Hemarajata

Pathology and Laboratory Medicine, University of California, Los Angeles, Los Angeles, California, USA

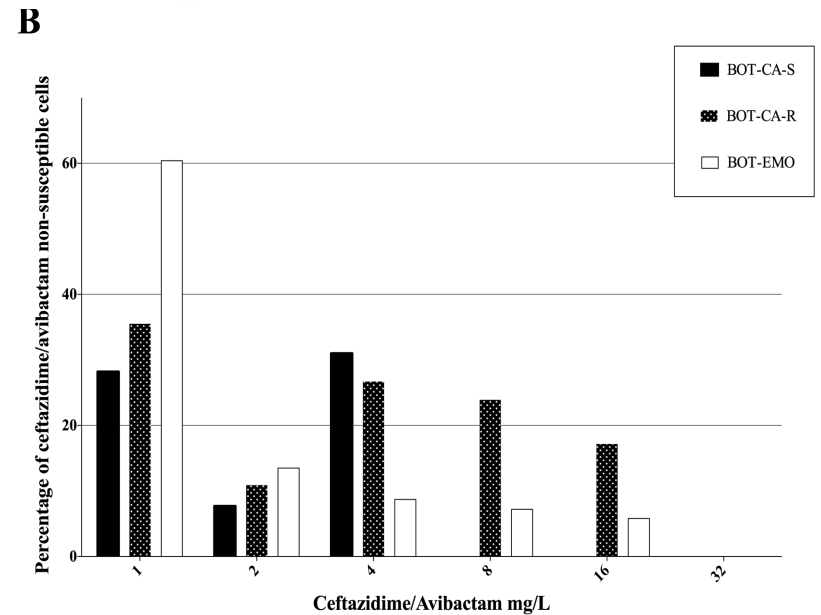
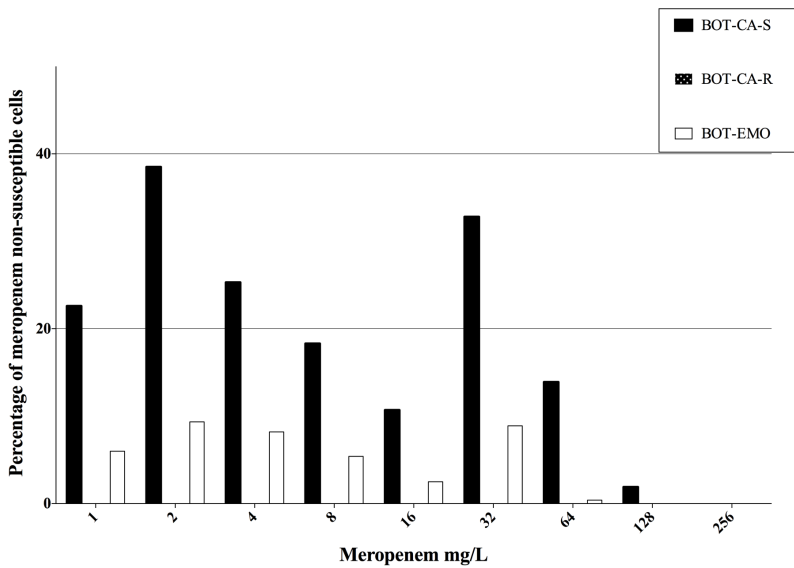
KEYWORDS ceftazidime-avibactam, KPC, OmpK35, OmpK36, resistance

AAC 2017; 61: e00537-17.

Heteroresistance to ceftazidime-avibactam



Gaibani P et al.
JAC 2018

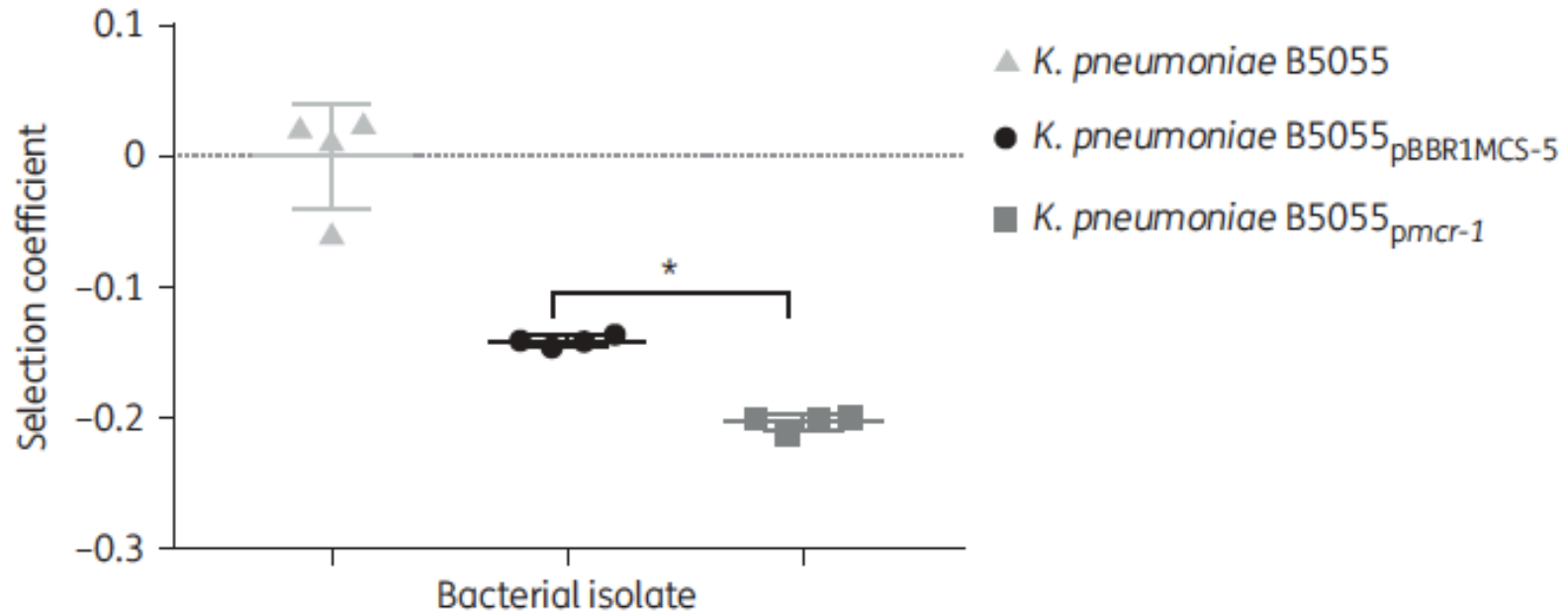


Mobile colistin resistance not the most common, but gets most attention...

MCR-type	Country	Species	Source
<i>mcr-1</i>	China	<i>E. coli</i>	Animal
<i>mcr-1.2</i>	Italy	<i>K. pneumoniae</i>	Human
<i>mcr-1.3</i>	China	<i>Salmonella</i>	Animal
<i>mcr-2</i>	Belgium	<i>E. coli</i>	Animal
<i>mcr-3</i>	China	<i>E. coli</i>	Animal
<i>mcr-4</i>	Italy	<i>Salmonella</i>	Animal
<i>mcr-5</i>	Germany	<i>Salmonella</i>	Animal

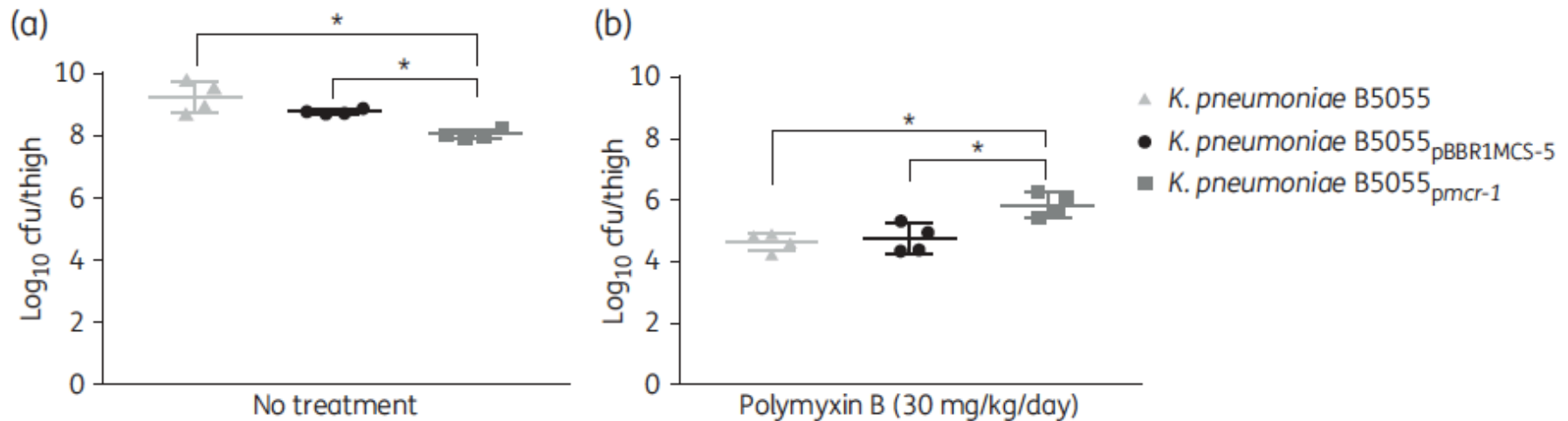
Liu et al. Lancet ID 2016; Xavier et al. Euro Surveill 2016;
Di Pilato et al. AAC 2016; Yin et al. mBio 2017;
Carattoli et al. Euro Surveill 2017; Borowiak et al. JAC 2017;
Lu et al. AAC 2017

Fitness cost of carrying *mcr-1*



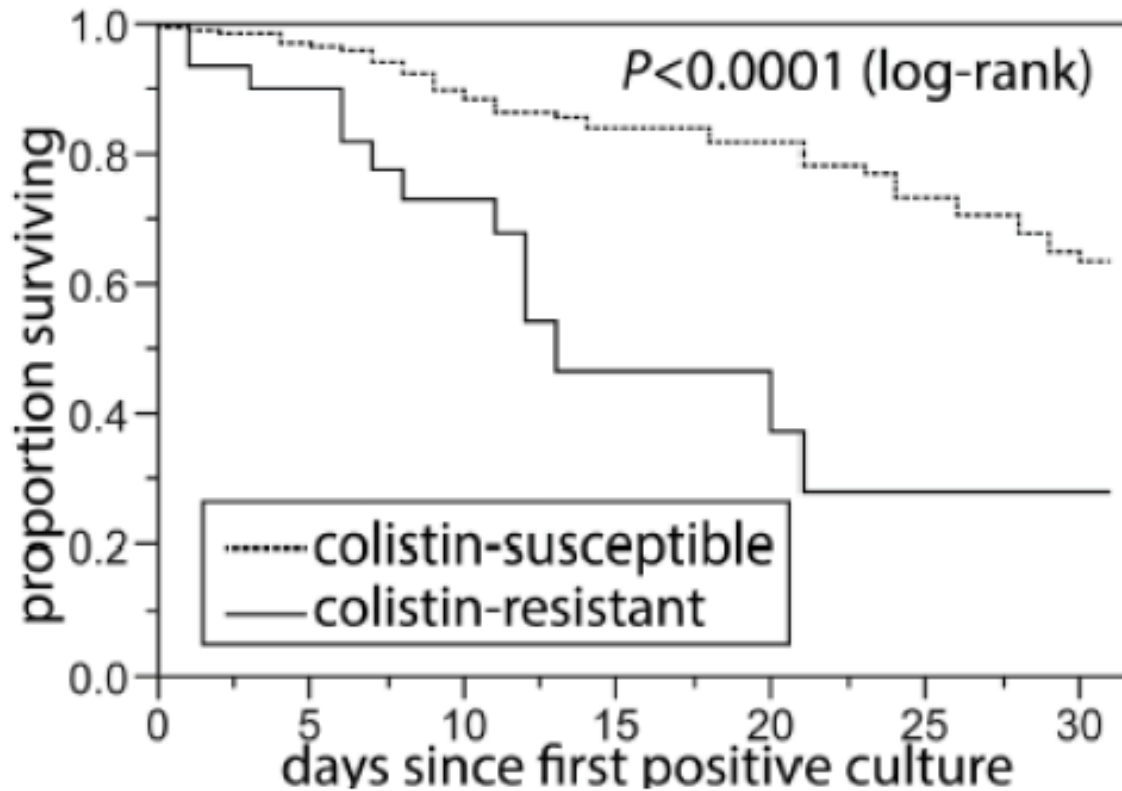
- In vitro competition assay (vs *E. coli* JW1)
- *K. pneumoniae* parental strain defined as selection coefficient zero
- Nang SC et al. JAC 2018

mcr-1 confers a selective advantage only under selective pressure with polymyxins



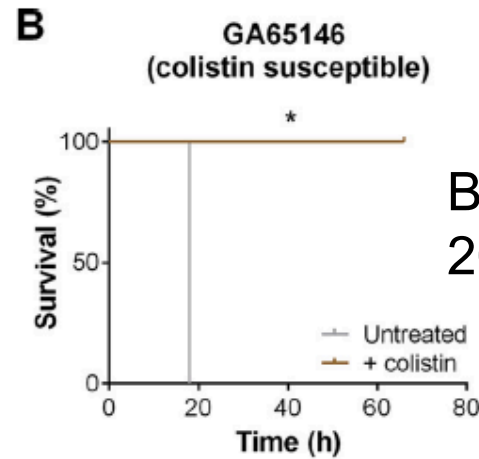
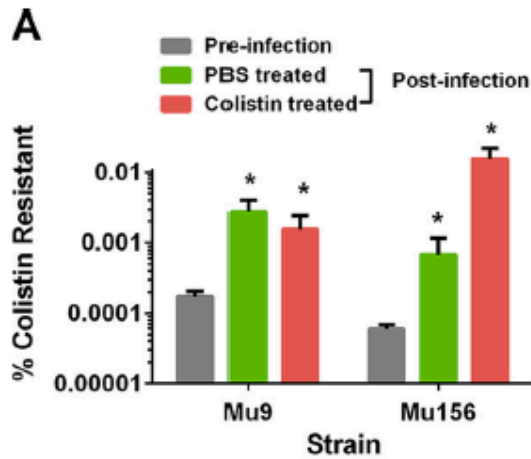
- Could indicate that e.g. carriage of *mcr-1* could have a low impact in individuals that are not exposed to polymyxins
- Consistent with the emergence of *mcr* in regions where polymyxin consumption is high

Colistin-resistant *K. pneumoniae* – survival in carbapenemase-producers

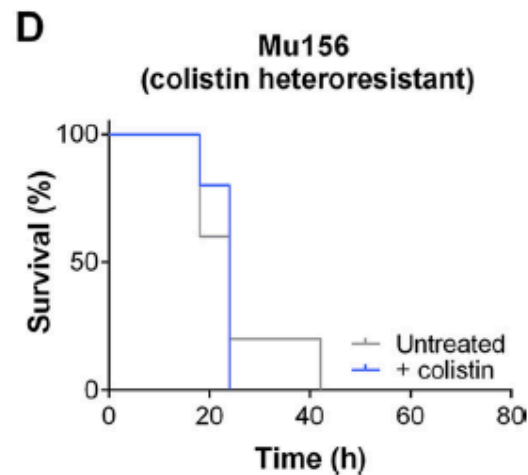
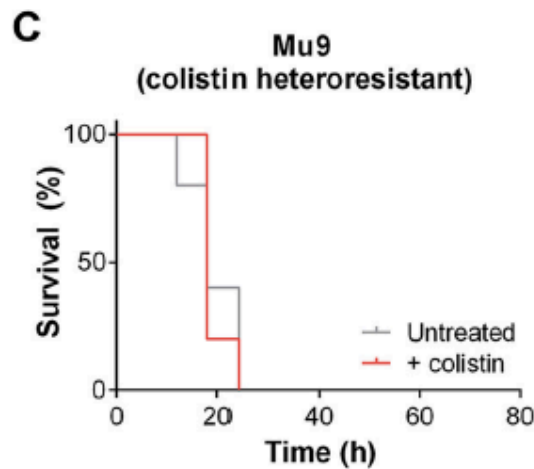


Rojas LR. CID 2017

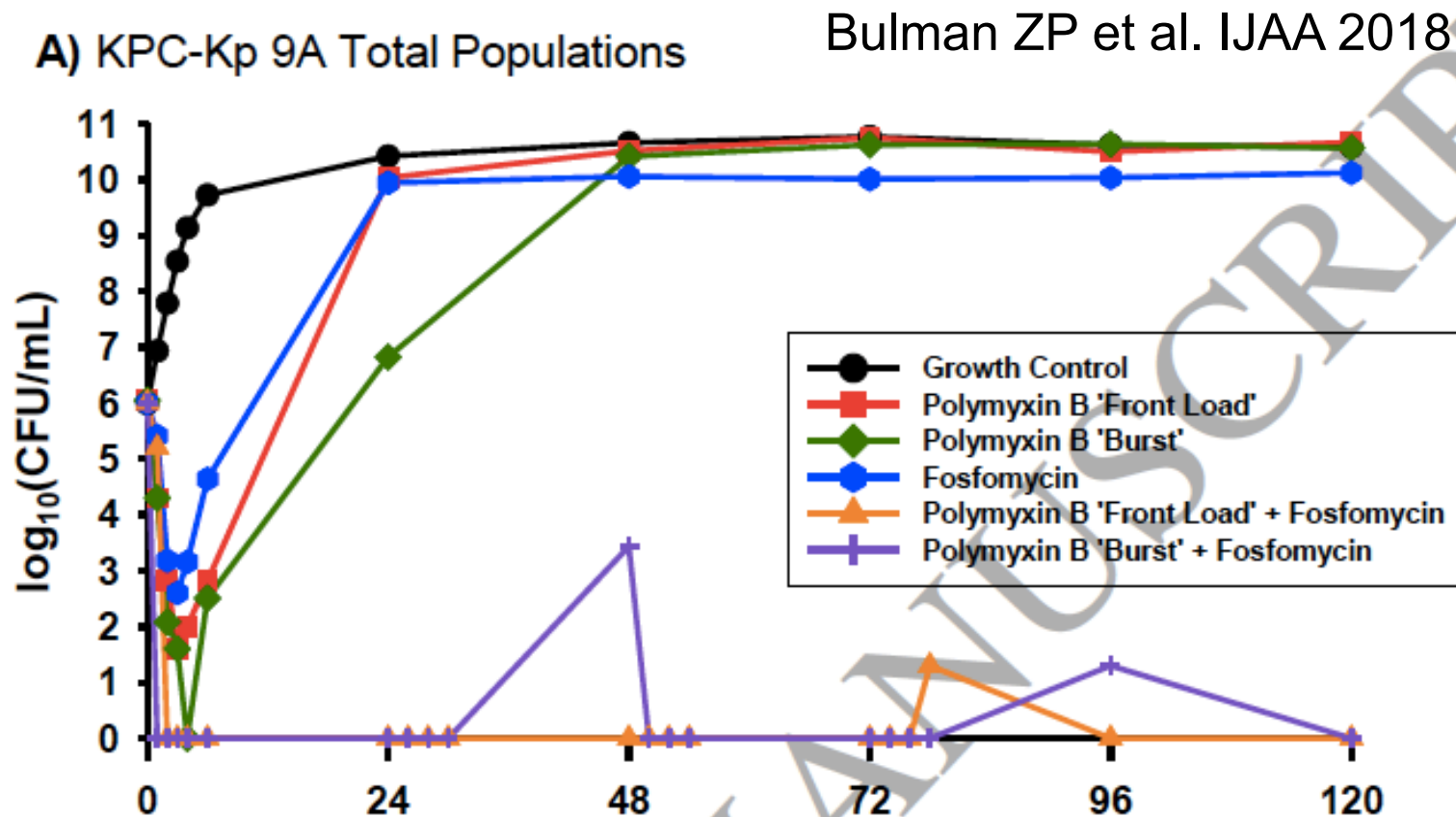
Colistin heteroresistance matters in the murine peritonitis model



Band VI, et al. mBio
2018; 9:e02448-17.



Hollow fiber: polymyxin B+fosfomycin prevents regrowth

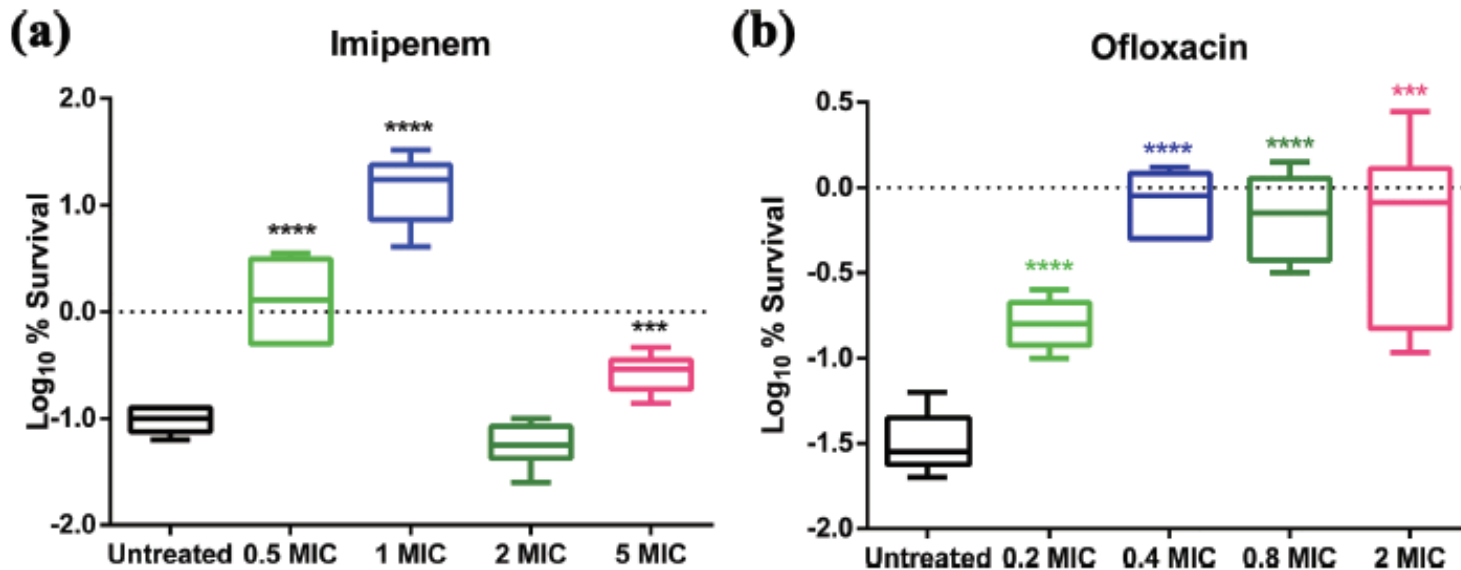


Pan-resistant *K. pneumoniae*

Class and antimicrobial(s)	MIC(s) (µg/ml)	Interpretation	Associated resistance gene(s)
Aminoglycoside			
Amikacin	>64	R	<i>aacA4, rmtC</i>
Gentamicin	>16	R	<i>aacA4, rmtC</i>
Tobramycin	>16	R	<i>aacA4, rmtC</i>
Beta-lactam			
Ampicillin	>32	R	<i>bla_{CTX-M-15s}, bla_{SHV-2B}, bla_{CMY-6}, bla_{NDM-1}</i>
Aztreonam	>64	R	<i>bla_{CTX-M-15s}, bla_{CMY-6}</i>
Cefazolin	>8	R	<i>bla_{CTX-M-15s}, bla_{SHV-2B}, bla_{CMY-6}, bla_{NDM-1}</i>
Cefepime	>32	R	<i>bla_{CTX-M-15s}, bla_{NDM-1}</i>
Cefotaxime	>64	R	<i>bla_{CTX-M-15s}, bla_{CMY-6}, bla_{NDM-1}</i>
Cefotaxime-clavulanic acid	>32/4	ND	<i>bla_{CMY-6}, bla_{NDM-1}</i>
Cefoxitin	>16	R	<i>bla_{CMY-6}, bla_{NDM-1}</i>
Ceftazidime	>128	R	<i>bla_{CTX-M-15s}, bla_{CMY-6}, bla_{NDM-1}</i>
Ceftazidime-avibactam ^b	>16/4	R	<i>bla_{NDM-1}</i>
Ceftazidime-clavulanic acid	>64/4	ND	<i>bla_{CMY-6}, bla_{NDM-1}</i>
Ceftriaxone	>32	R	<i>bla_{CTX-M-15s}, bla_{CMY-6}, bla_{NDM-1}</i>
Doripenem	>8	R	<i>bla_{NDM-1}</i>
Ertapenem	>8	R	<i>bla_{NDM-1}</i>
Imipenem	32	R	<i>bla_{NDM-1}</i>
Meropenem	>8	R	<i>bla_{NDM-1}</i>
Piperacillin-tazobactam	>128/4	R	<i>bla_{NDM-1}</i>
Chloramphenicol			
Chloramphenicol	>16	R	Truncated <i>ramR</i>
Fluoroquinolone			
Ciprofloxacin	>8	R	<i>oqxA, oqxB, gyrA</i> and <i>parC</i> mutations, truncated <i>ramR</i>
Levofloxacin	>8	R	<i>oqxA, oqxB, gyrA</i> and <i>parC</i> mutations, truncated <i>ramR</i>
Fosfomycin			
Fosfomycin	32 ^c , 16 ^d	ND	<i>fosA</i>
Polymyxin			
Colistin	>8	NWT	Disrupted <i>mgrB</i>
Polymyxin-B	>8	NWT	Disrupted <i>mgrB</i>
Sulfonamide			
Trimethoprim-sulfamethoxazole	8/152	R	<i>sul1</i>
Tetracycline			
Tetracycline	>32	R	<i>tet(A)</i> , truncated <i>ramR</i>
Tigecycline ^b	4	I	Truncated <i>ramR</i>
Macrolide			
Not included in AST panel	Na	Na	<i>mph(A)</i>

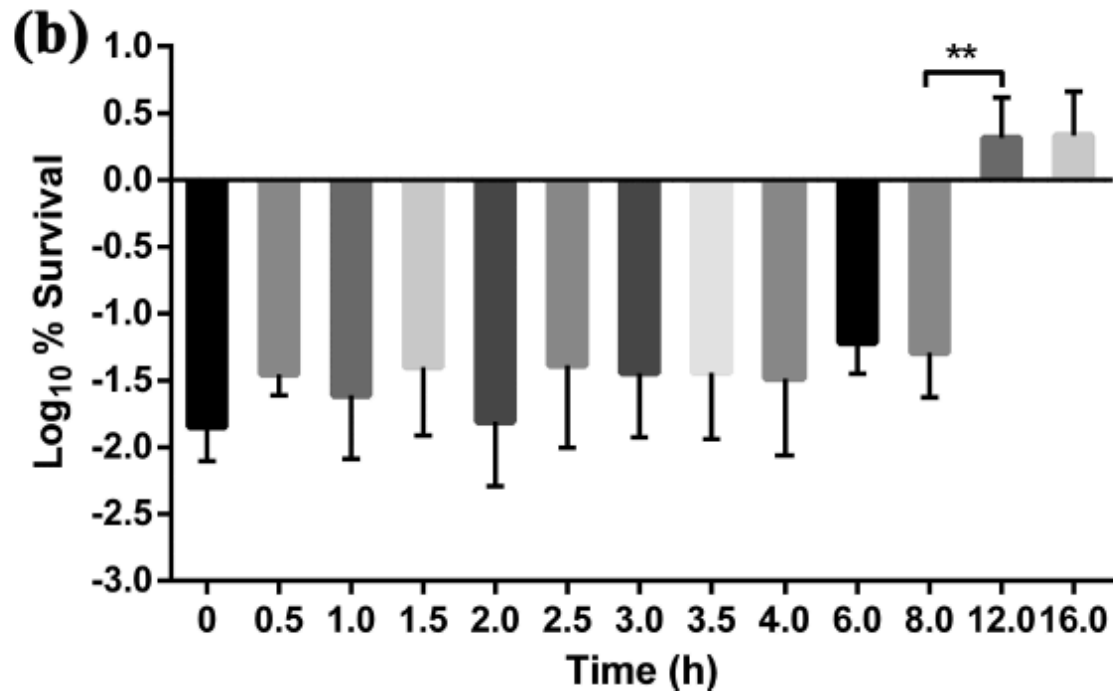
de Man TJB, et al.
mBio 2018; 9:
e00440-18.

Persister cells are commonly formed with *K. pneumoniae* grown in exponential phase



- Persister cells could play a role in the frequent development of resistance
- Li Y et al. J Medical Microbiol. 2018; 67: 273-281

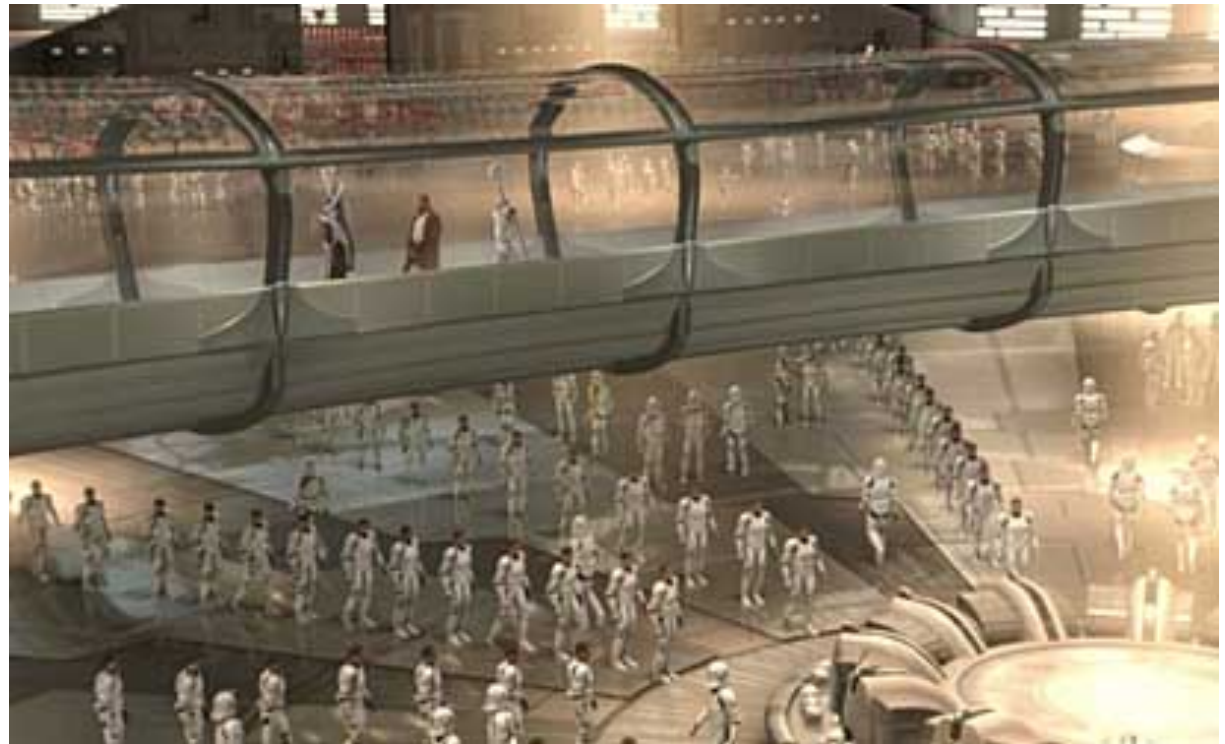
Persister cells in stationary phase can even survive in 100-fold MIC concentrations



- Number of hours from subcultivation
 - Statistically significant when comparing late exponential phase and stationary phase (could be of importance in subacute infections/colonization)
-

Bacterial clones

What is a clone?



History of phylogenetic typing

- Multilocus enzyme electrophoresis (MLEE)
 - Identifies variants of the gene products of 10–20 housekeeping genes (genes encoding basic metabolic functions)
 - Electrophoresis of cell extracts on starch gels, followed by detection using specific enzyme stains
 - Problem: interlaboratory variation
- 1998 introduction of multilocus sequence typing (MLST)
 - Increased resolution, few loci needed
 - Exportable data and international databases
 - *N. meningitidis* and *S. pneumoniae* both available in 1998
- Difficult to visualize population structures with trees as the number of sequences increase

Clonal expansion – MLST definition (strict)

Isolate	Gene A	Gene B	Gene C	Gene D	Gene E	Gene F	Gene G
1	1	1	1	1	1	1	1
2	1	1	1	1	1	1	2
3	1	1	1	1	1	2	1
4	1	1	1	1	2	2	2

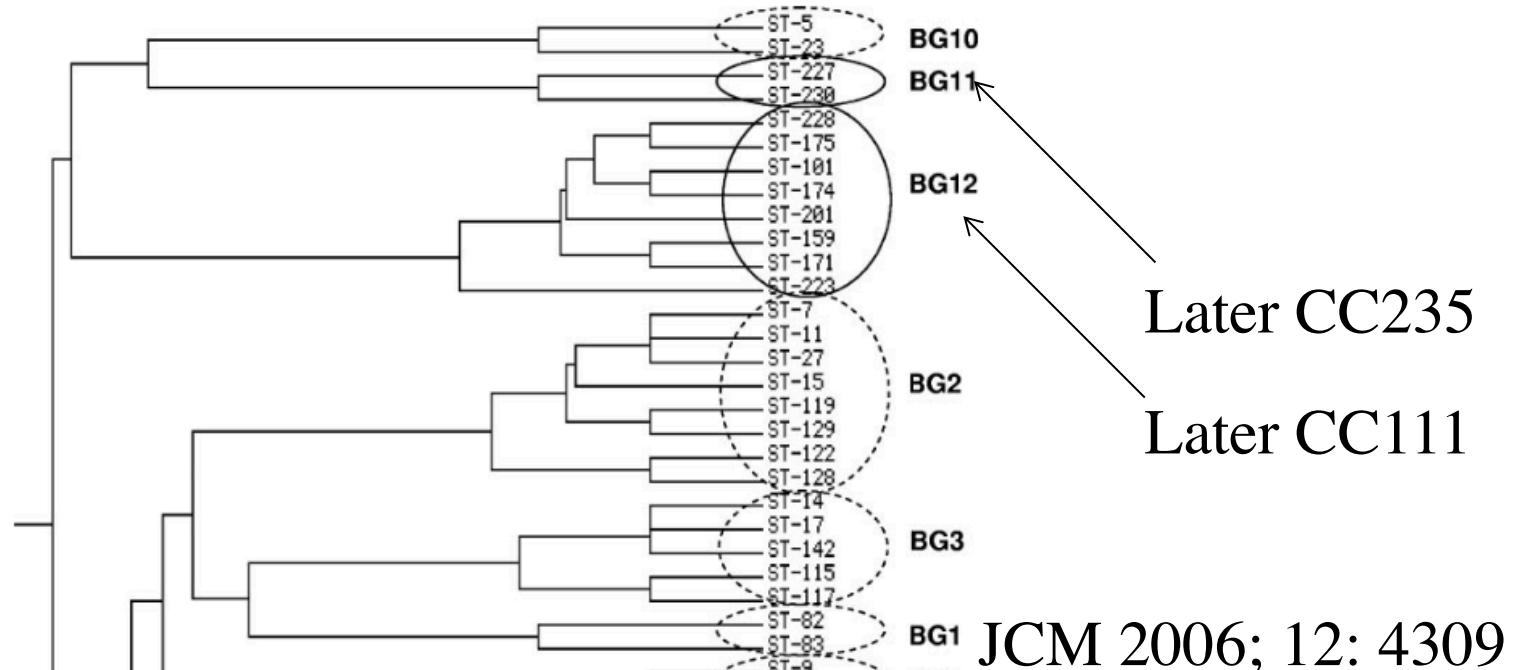
High-Risk Clones (HiRC)

- Associated with resistance of great clinical importance
- AND
- High ability to spread in health-care institutions
- OR
- High ability to confer invasive disease
- OR
- High ability to colonize individuals for long time periods
-
- Perhaps a better term: epidemic clone
-

Some early work on epidemic clones

Establishing Clonal Relationships between VIM-1-Like Metallo- β -Lactamase-Producing *Pseudomonas aeruginosa* Strains from Four European Countries by Multilocus Sequence Typing[∇]

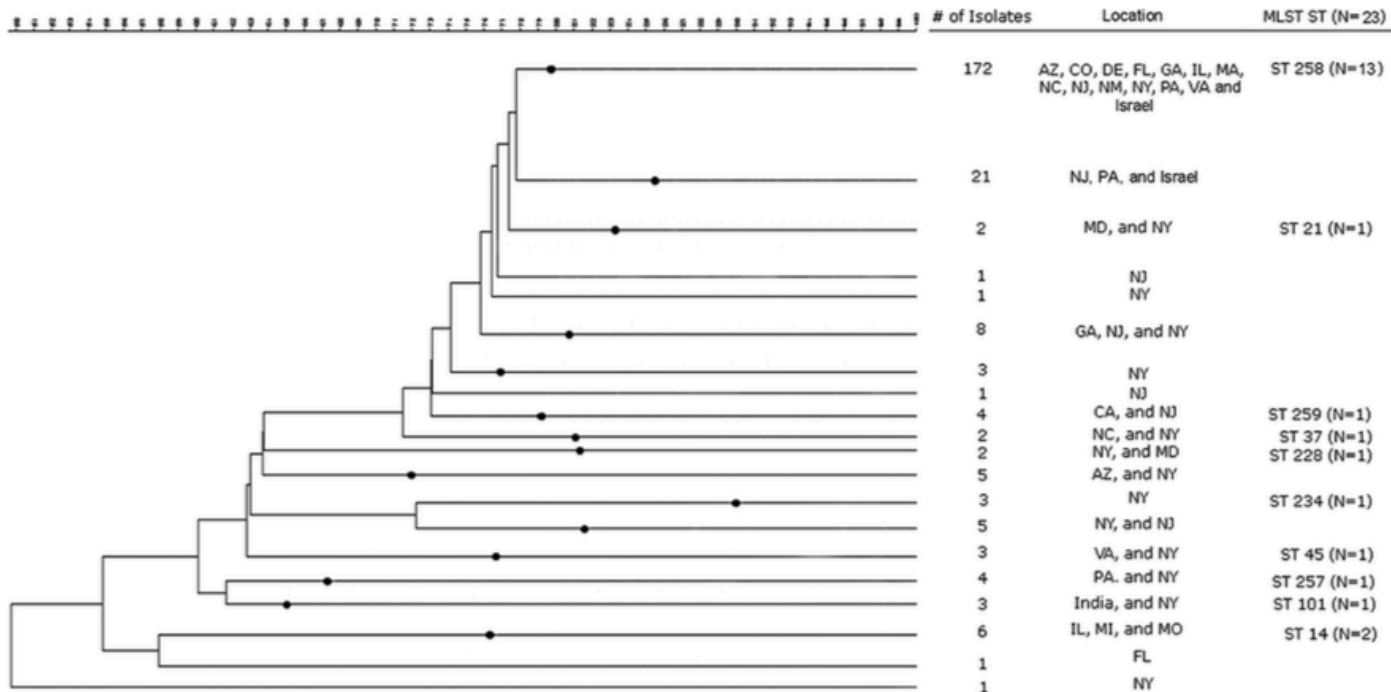
Christian G. Giske,^{1†*} Balázs Libisch,^{2†*} Céline Colimon,^{3‡} Effie Scoulica,⁴ Laura Pagani,⁵
Miklós Füzi,² Göran Kronvall,¹ and Gian Maria Rossolini³



Early work in KPC-producing *K. pneumoniae*

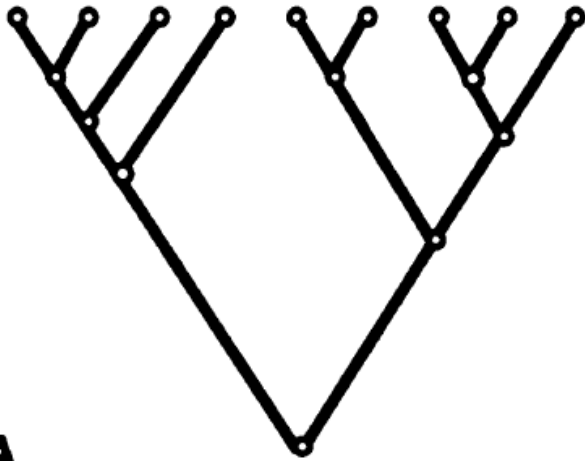
Molecular Epidemiology of KPC-Producing *Klebsiella pneumoniae* Isolates in the United States: Clonal Expansion of Multilocus Sequence Type 258[▽] AAC 2009; 8: 3365

Brandon Kitchel,^{1*} J. Kamile Rasheed,¹ Jean B. Patel,¹ Arjun Srinivasan,¹ Shiri Navon-Venezia,² Yehuda Carmeli,² Alma Brolund,³ and Christian G. Giske³



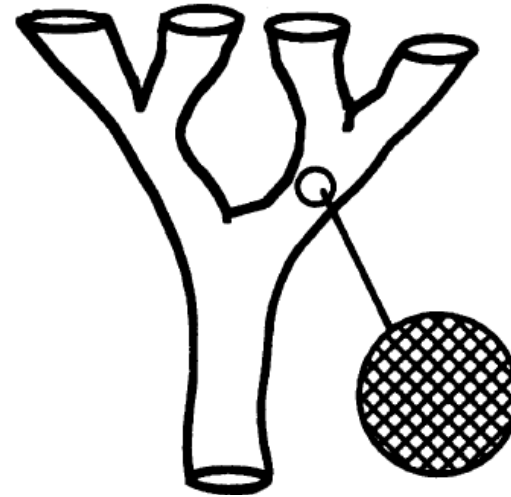
PFGE-
database:
n=248

Population structures



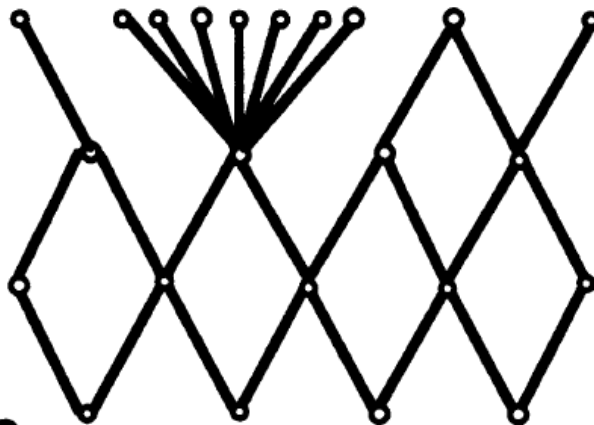
A

Clonal, no recombination



B

Recombination within main branches

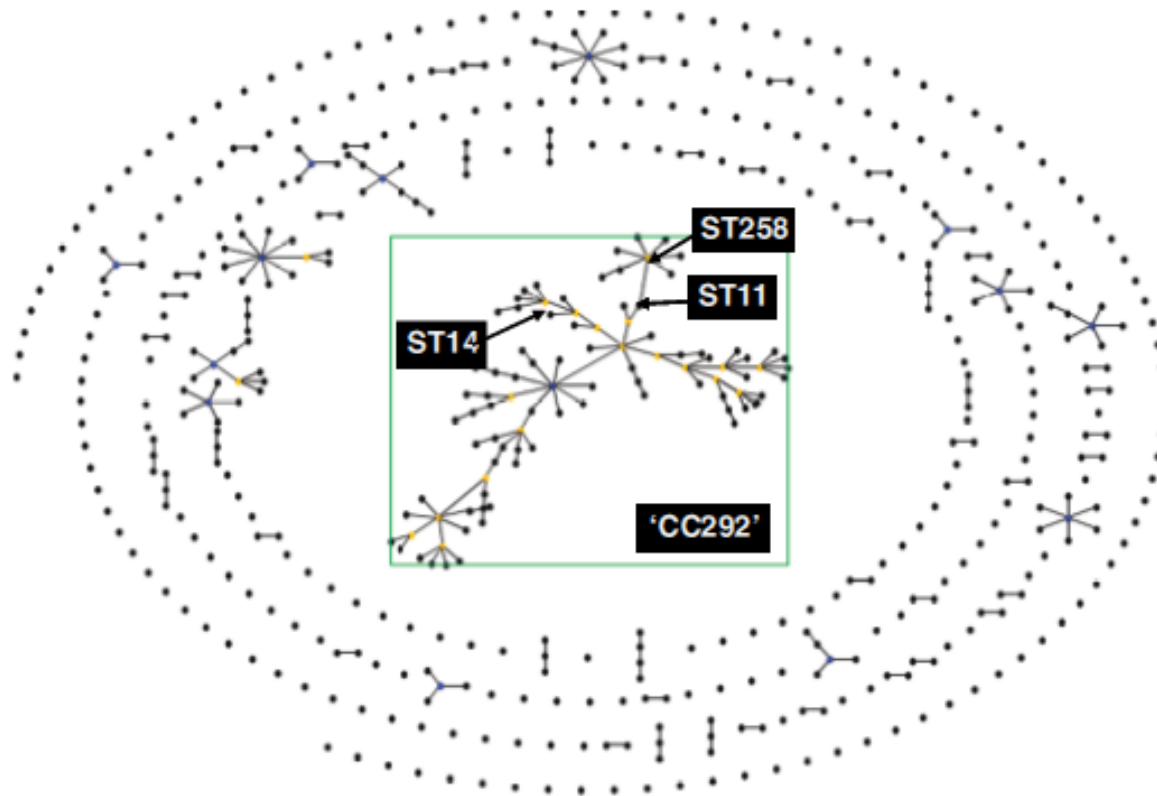


C

Panmictic/epidemic

Smith JM. PNAS 1993

Population snapshot *K. pneumoniae*

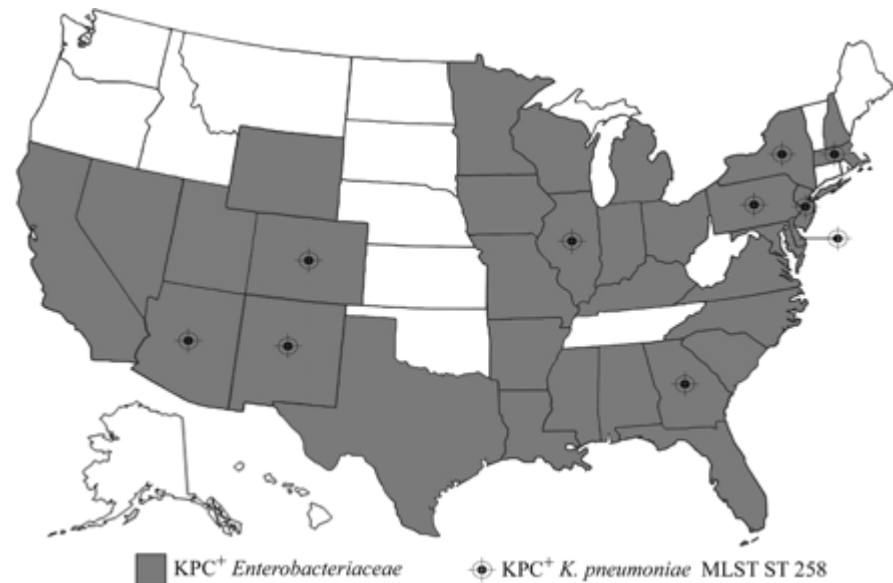


Woodford et al. FEMS Microbiol Lett 2011

K. pneumoniae ST258

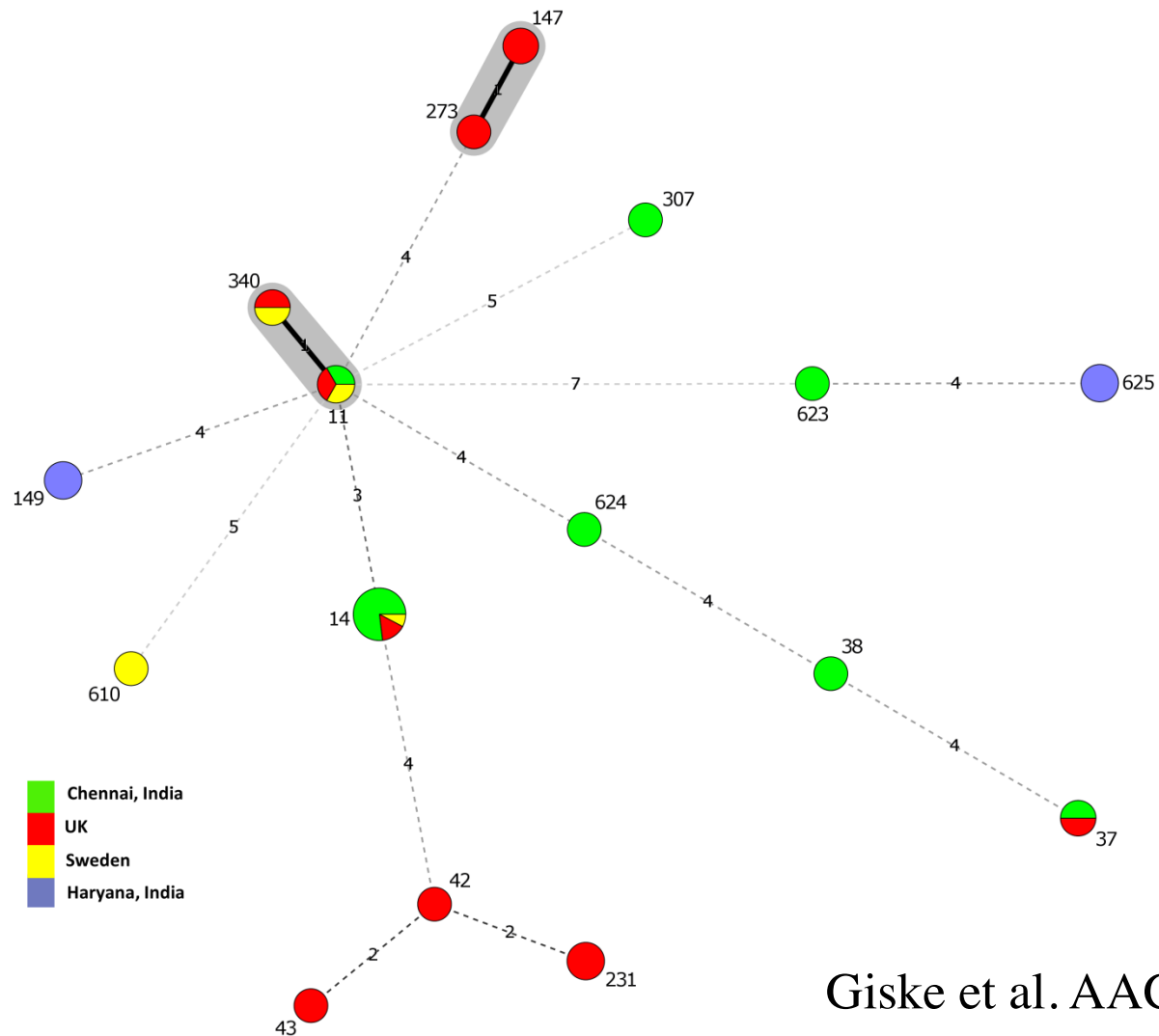


- Clone associated with the spread of the carbapenemase KPC
- Initially described in USA, then in most countries where KPC has been detected
- Responsible for 70% of the cases of KPC in the US and most cases in Israel
- Also capable of causing invasive disease



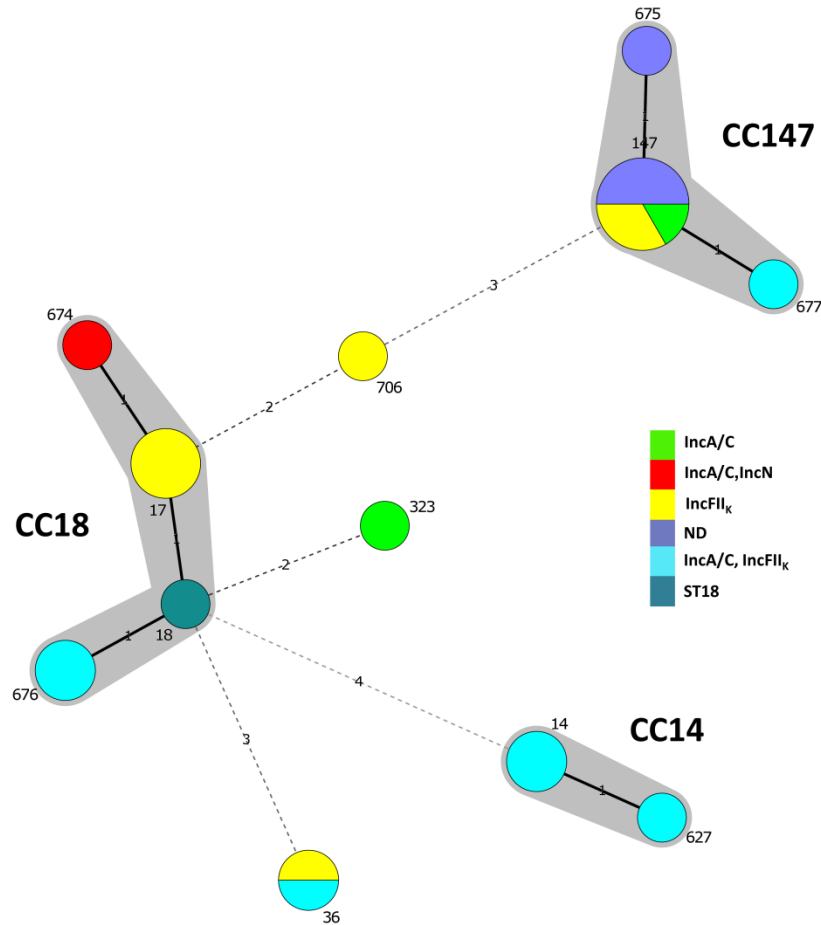
Kitchel et al. AAC 2009

Sequence types in NDM-produces

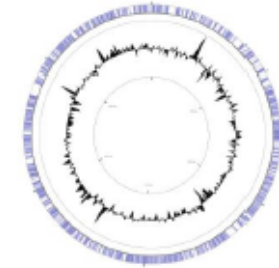
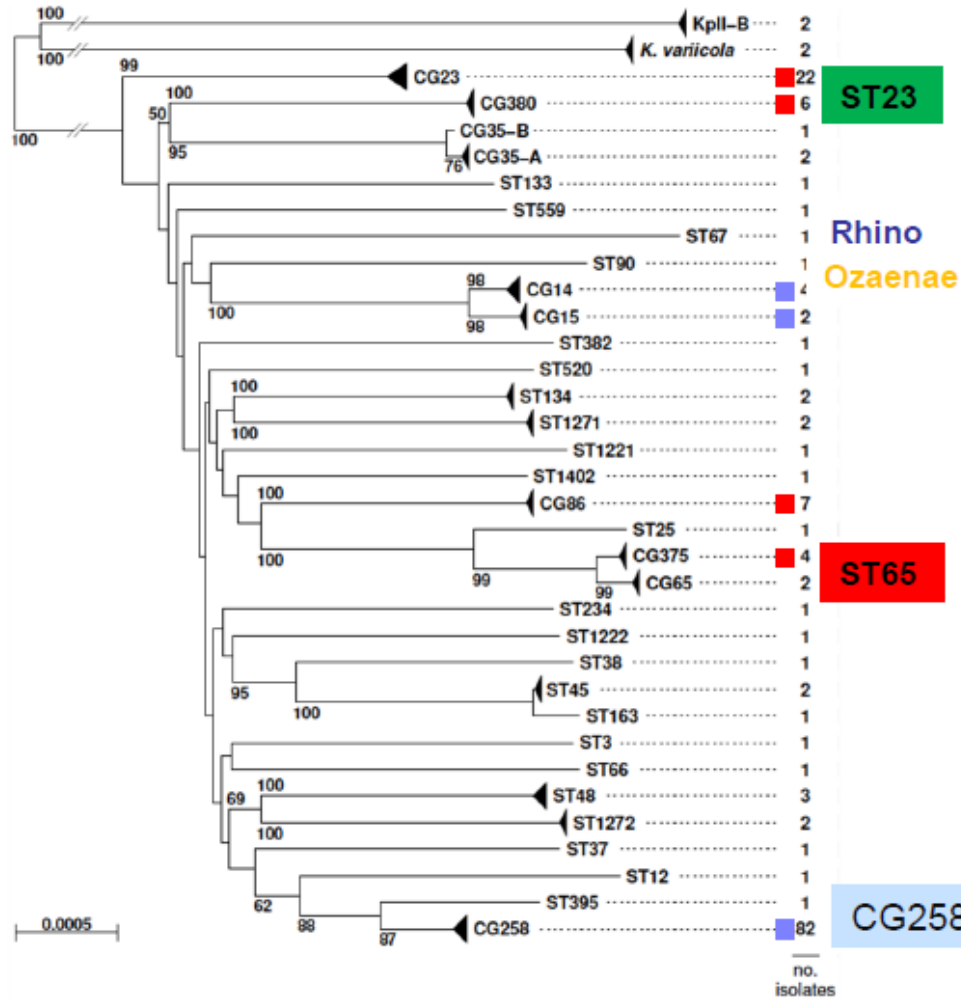


Giske et al. AAC 2012

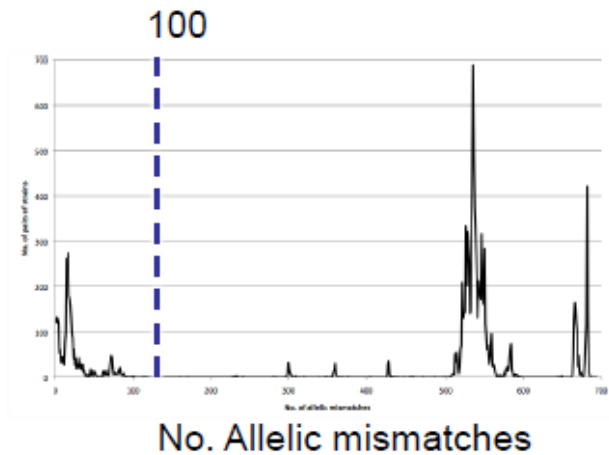
Sequence type in VIM-producers



Correlation cgMLST and MLST



694 core genes



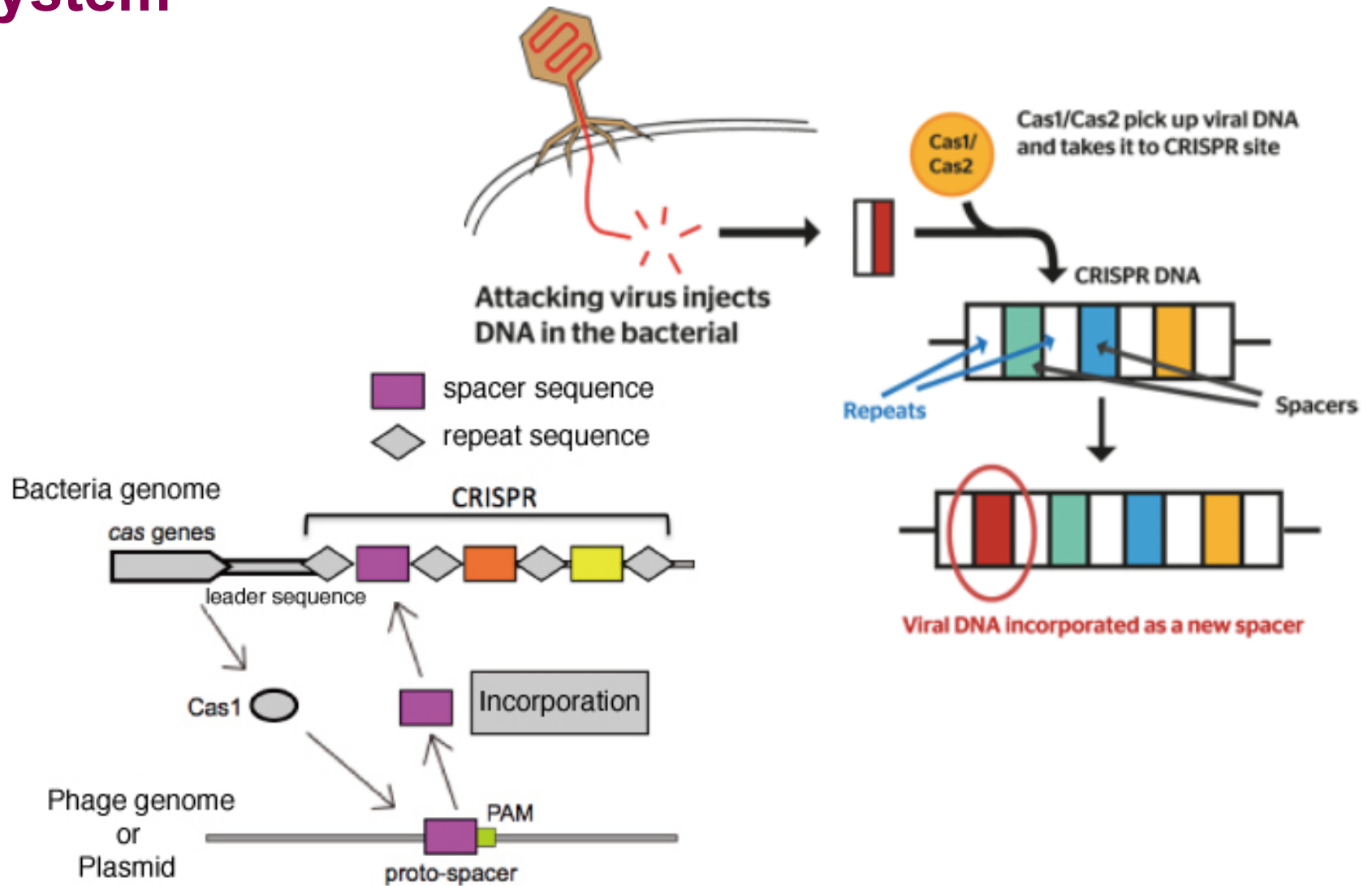
Epidemic clones of *K. pneumoniae*

Searching for differences between epidemic and non-epidemic clones in the genome

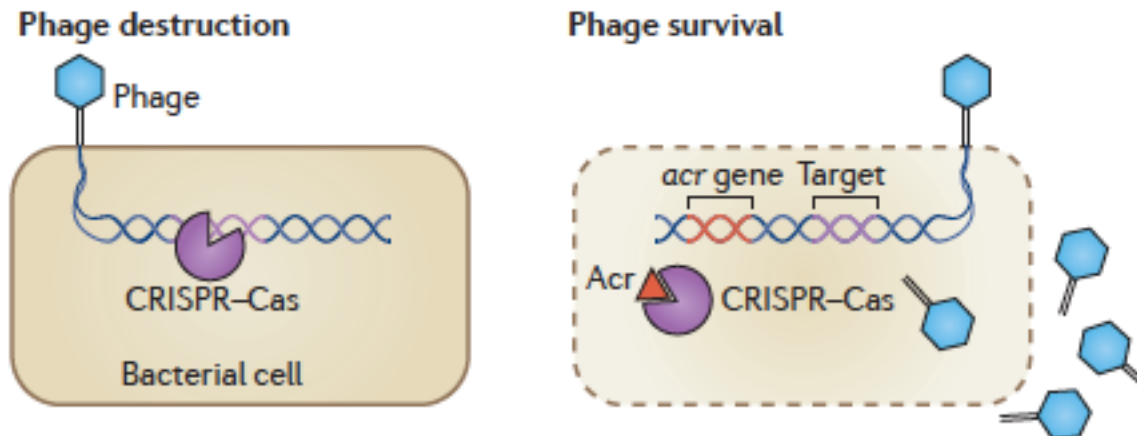
Feature	Difference epidemic vs non-epidemic
Virulence genes	Few virulence genes in both groups
Prophages	No quantitative difference between groups
CRISPR-Cas	Higher occurrence in epidemic ($p < 0.01$)

Örmälä-Odegrip A et al. Manuscript

CRISPR-Cas: the bacterial immune system



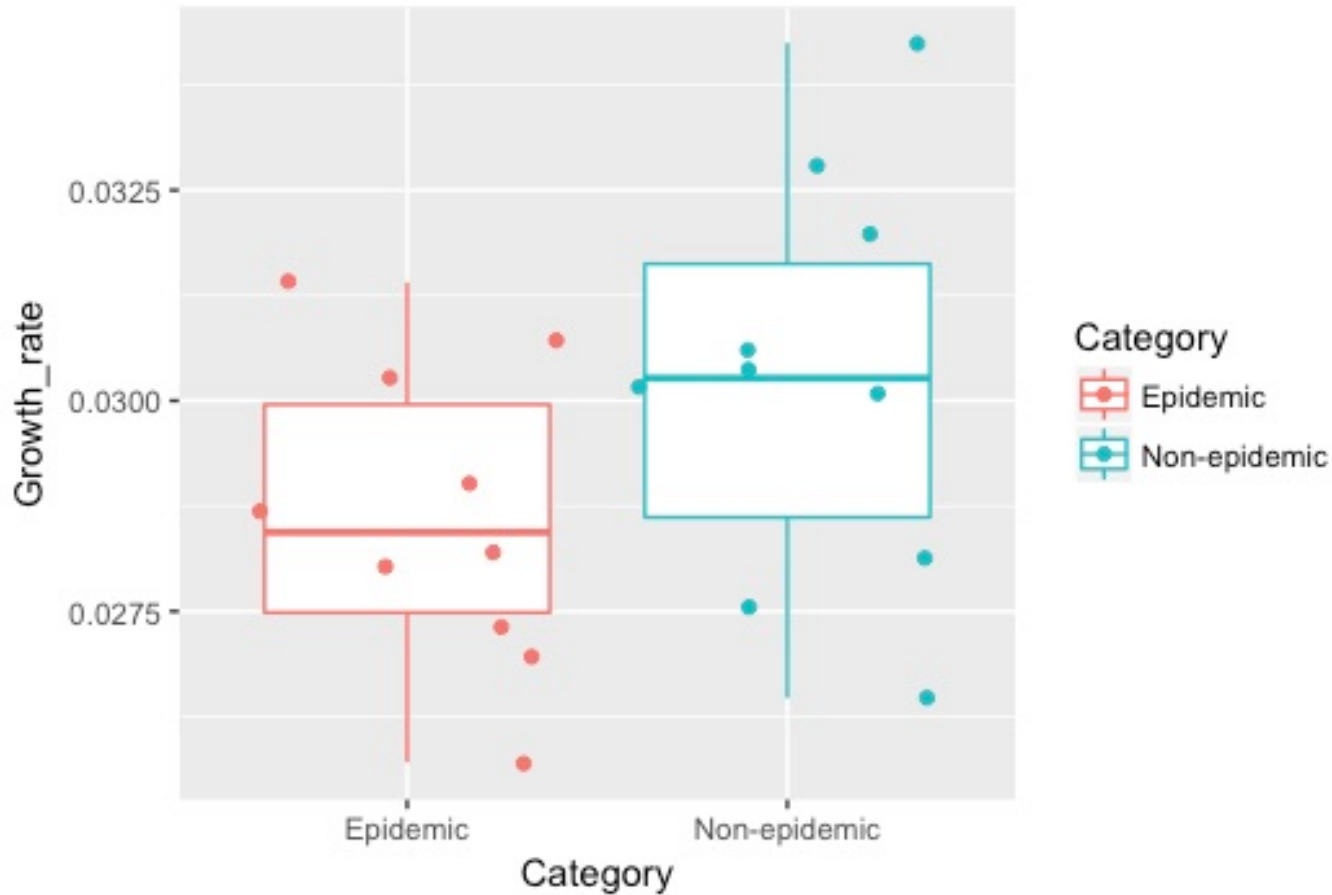
Anti-CRISPR: phages and mobile elements evade CRISPR-Cas immunity



Pawluk A. Nat Rev Micro 2017

Growth rate (Bioscreen)

Growth rate, bioscreen



GLM (ANOVA)

F =8.406

df= 1

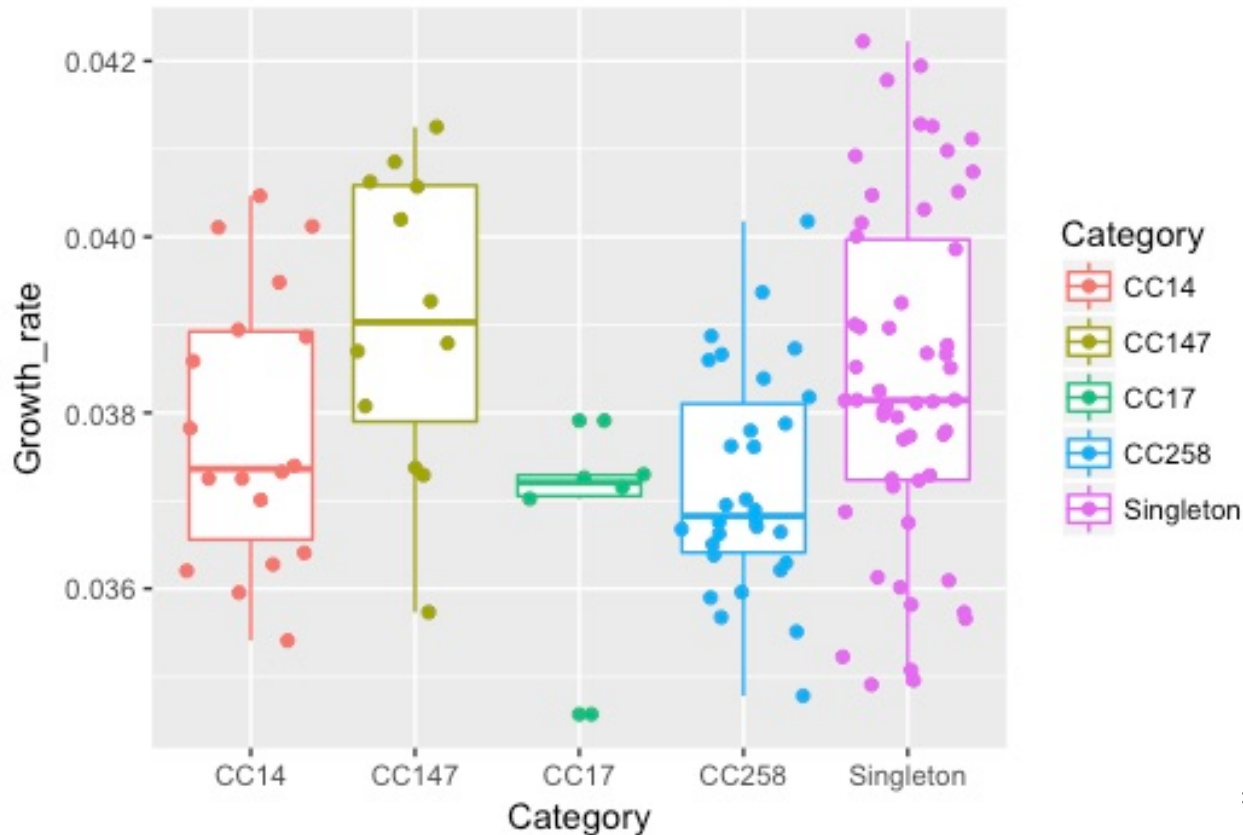
Sig.= 0.05 level

Student T-test

Sig.= 0.005 level

Non-epidemic isolates
grow faster

Growth rate per CC



GLM (ANOVA)

F =4.326

df= 4

Sig.= 0.003

Post Hoc *

CC258 < CC147,
Singleton

CC147 > CC258,
CC17

Singleton > CC258,
CC17

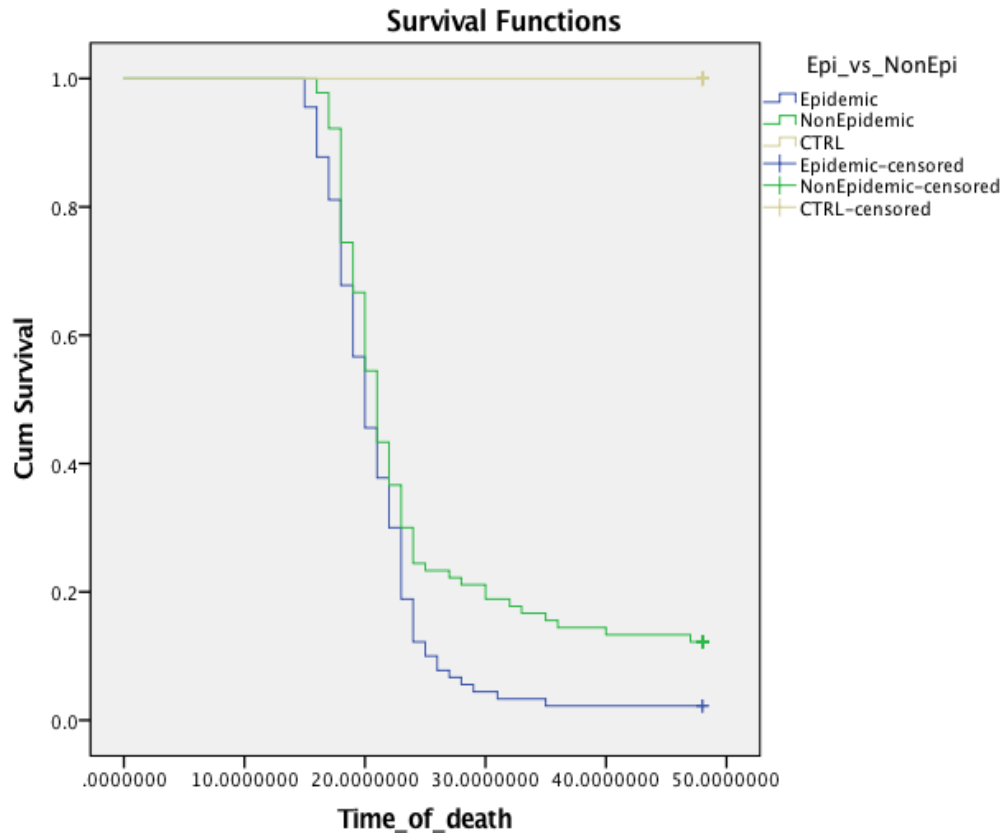
* The mean difference is
significant at 0.05 level

Virulence (*Galleria mellonella*)

Galleria mellonella

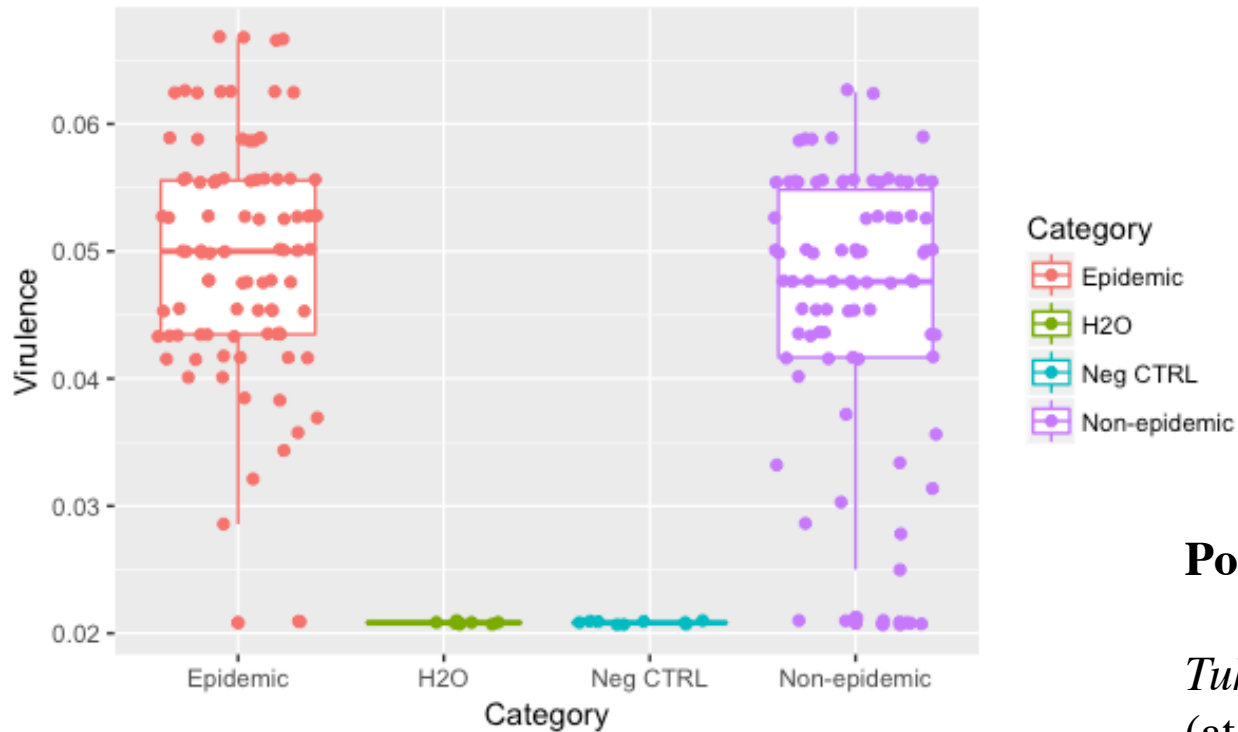


Survival epidemic vs non-epidemic



Overall Comparisons			
	Chi-Square	df	Sig.
Log Rank (Mantel-Cox)	7.536	1	.006
Breslow (Generalized Wilcoxon)	4.291	1	.038

Virulence: epidemic > non-epidemic



GLM (Epi vs Non-epi)

df= 1

F= 9.099

Sign.= 0.003

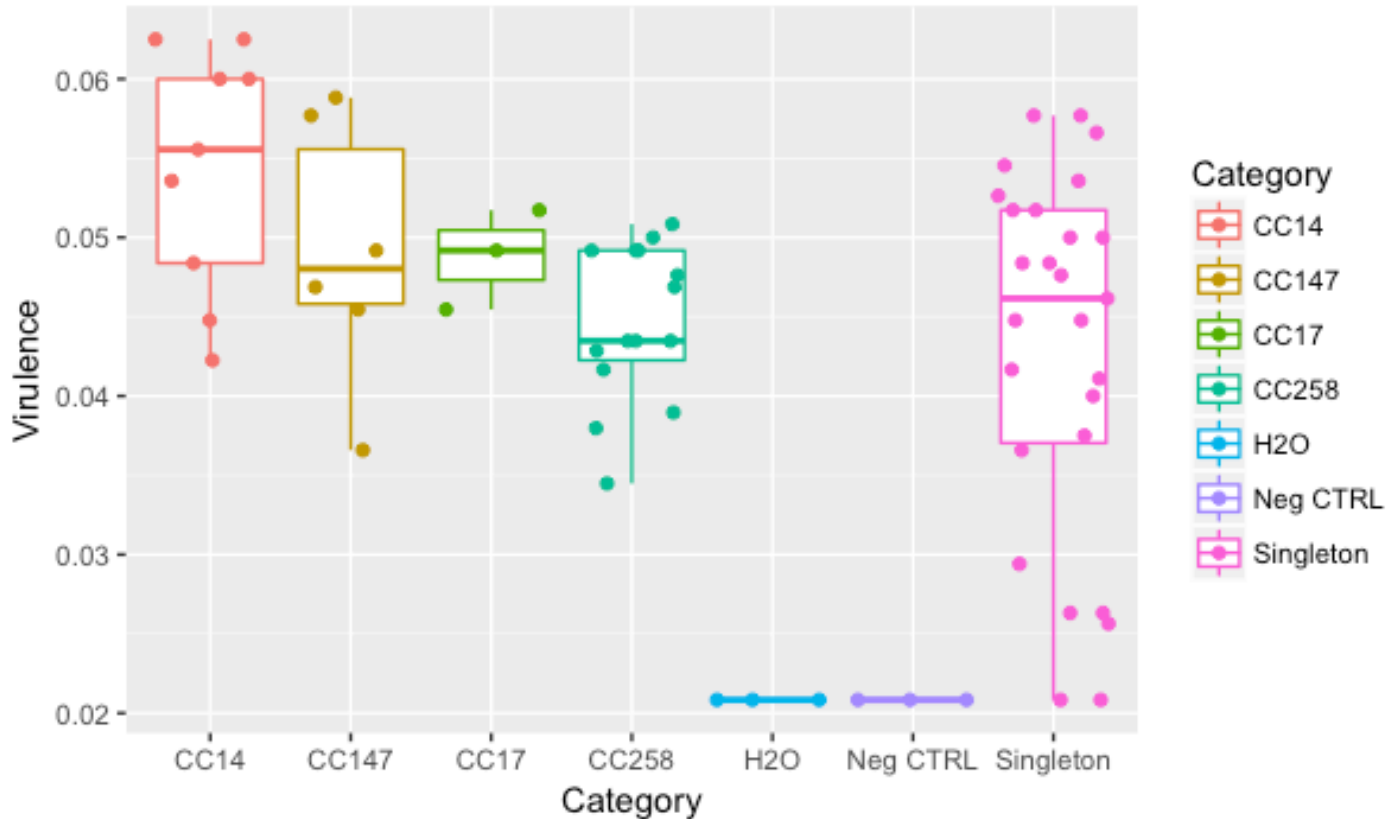
Poect Hoc: multiple comparison

Tukey: Epidemic > Non-epidemic
(at sig 0.05)

LSD: Epidemic > Non-epidemic
(at sig 0.02)

Mean difference is significant

Virulence vs clonal complex



GLM (Clonal Complex)

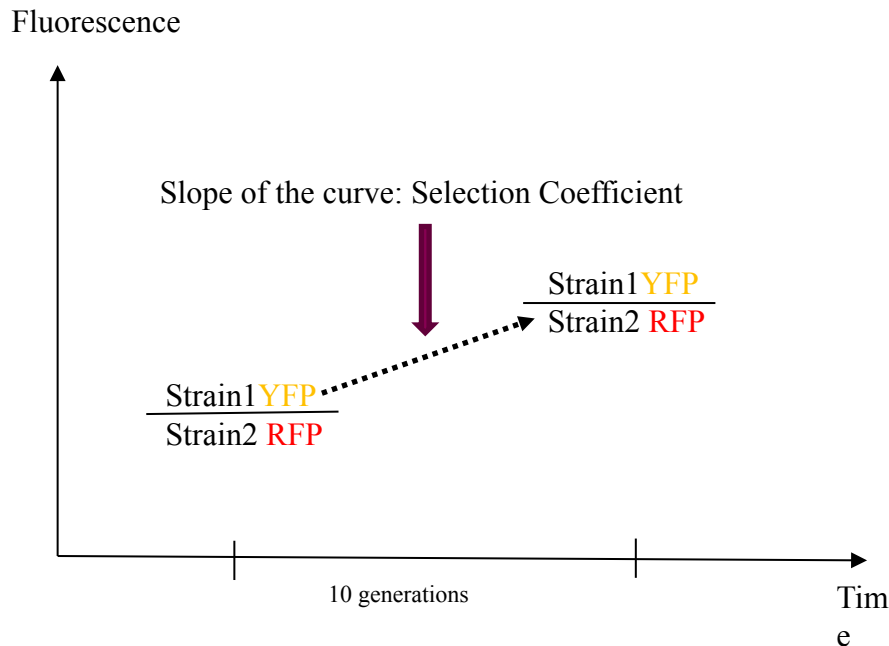
df= 4

F= 2.876

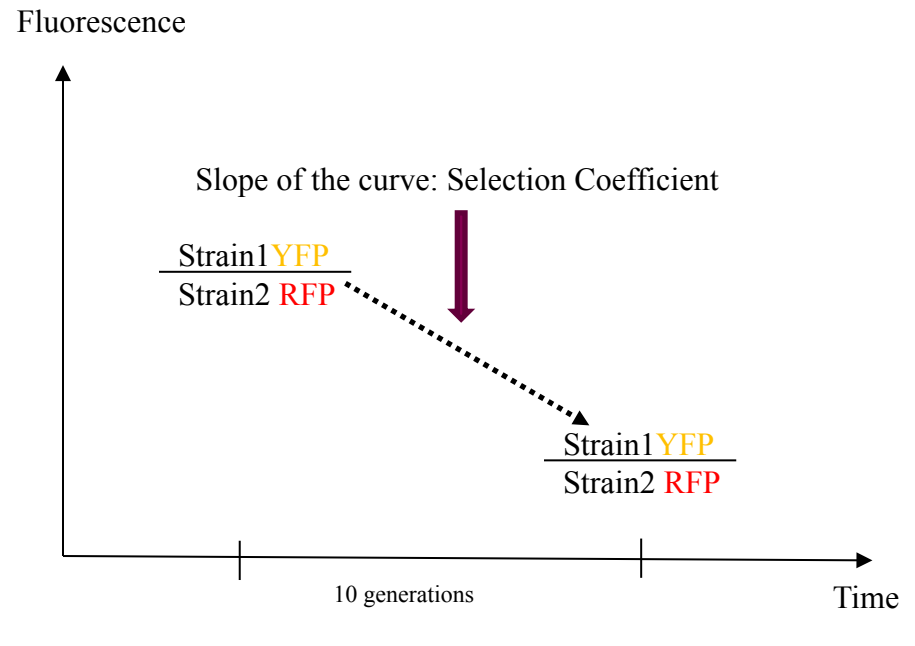
Sign.= .031

Liquid competitions (MACS)

Competition experiments: magnetic-activated cell sorting (MACS)



Positive slope → (+) S-value: Strain 1 wins over Strain 2

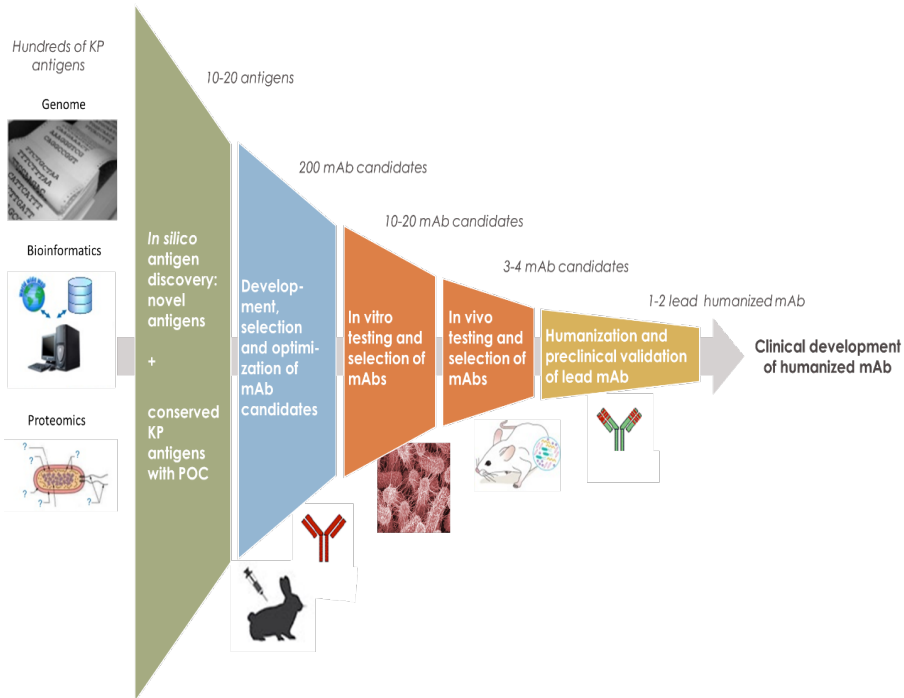


Negative slope → (-) S-value: Strain 1 loses against Strain 2

Preliminary assessment: non-epidemic strains seem to win over epidemic strains in majority of cases

Novel treatment concepts

Novel treatment concepts



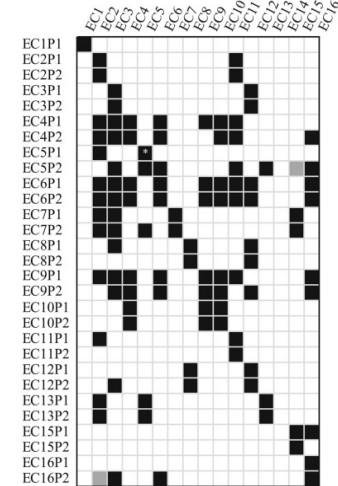
1. Sample collection



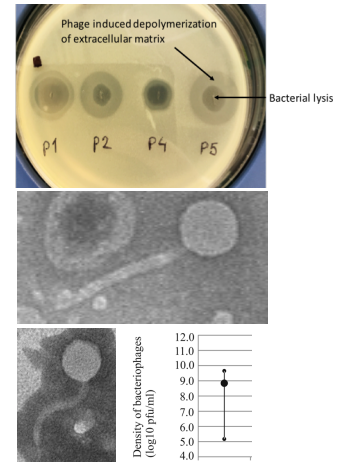
2. Phage enrichment / isolation



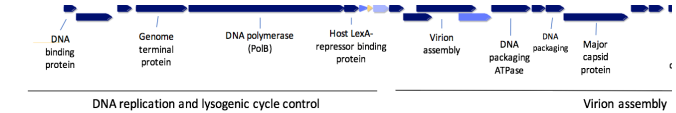
3. Host range



4. Phenotypic characterization



5. Genomic analysis



Conclusions

- In some regions *K. pneumoniae* infections are observed in multimorbid individuals
 - Successful (or epidemic) clones are responsible for a major share of dissemination of antimicrobial resistance
 - Noteworthy among resistance mechanisms are particularly the carbapenemases
 - Heteroresistance may be important in resistance to several drug classes – possible to suppress with combination therapy?
 - Molecular typing tools have helped us define important clones
 - The factors determining why some clones are epidemic are still largely unknown – detecting them only a first step
 - Novel treatment concepts: phages and antibodies
-